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(54) Title: HUMAN BRAIN SODIUM-DEPENDENT INORGANIC PHOSPHATE COTRANSPORTER

(57) Abstract

This invention describes a novel human brain Na+-dependent inorganic phosphate cotransporter, designated the hBNPI protein. This invention also encompasses nucleic acids encoding this protein, or a fragment thereof, as well as methods employing this protein and the nucleic acid compounds.

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HUMAN BRAIN SODIUM-DEPENDENT INORGANIC PHOSPHATE COTRANSPORTER

Inorganic phosphate (Pi), a charged anion, 5 essential to bioenergetics, metabolic regulation, and bone and membrane structure. It is well known that Pi homeostasis in the body depends primarily on mechanisms that govern the renal excretion of P_i into the glomerular filtrate and its subsequent reabsorption against an electrochemical gradient 10 via brush-border epithelial cells located in the proximal tubule of the kidney [J. Bonjour and J. Caverzasio, Reviews in Physiological Pharmacology, 100:161-214 (1985); V.W. Dennis, Phosphate homeostasis, in HANDBOOK OF PHYSIOLOGY, (S. Shultz, ed. 1991) at pages 1785-1815.] This transepithelial transport of P_i is mediated, in part, by a transport system 15 which is driven by the transmembrane Na⁺ gradient across the microvilli brush border membrane. However, it remains largely unknown how cells transport and regulate necessary the intracellular concentrations of Pi, and the molecular events 20 underlying this system. Experiments using isolated kidney tubules or brush-border membranes have shown that Pi transport is rather complex, regulated not only by extracellular [Pi] but also by neurotransmitters such as catecholamines (for review see V.W.Dennis, supra), and by a variety of hormones and metabolic factors. Berndt and Knox, 25 "Renal Regulation of Phosphate Excretion", in, THE KIDNEY. PHYSIOLOGY AND PATHOPHYSIOLOGY, (D.W. Seldin and G. Giebisch, eds., 1991) at pages 1381-1396. Renal denervation, for example, decreases sodium and phosphate reabsorption. 30 Norepinephrine released from nerve endings in proximity to renal tubules acts on the proximal tubule to increase phosphate reabsorption. In studies of isolated tubules, however, dopamine is shown to inhibit phosphate and sodium transport in the rabbit proximal tubule. Furthermore, several 35 studies also show that depletion of extracellular Pi or increased circulating levels of parathyroid hormone alter the activity and expression of transporter molecules or both.

Several recent reports have demonstrated that Pi homeostasis significantly affects the central nervous system (CNS). Phosphate/calcium alterations in serum, for example, have been implicated in the etiology and pathogenesis of Alzheimer's diseases. Depletion of high energy phosphates 5 (phosphocreatine) and ATP is thought to be part of the final common pathway mediating excitotoxic neuronal cell death secondary to a wide variety of insults. Tight coupling between Pi transport and ATP production has been observed in many cells and tissues. Chronic P_i depletion in vivo is 10 associated with a significant reduction in the ATP content of polymorphonuclear leukocytes, platelets, and various tissues including kidney, heart, and skeletal muscle. A similar observation has been made in cultured peripheral vagal nerves. This reduction in intracellular ATP has been shown to 15 be a direct consequence of the decrease in intracellular Pi which occurs following Pi depletion. In addition to its possible role in ATP biosynthesis, several lines of evidence have suggested that Pi may be involved in neuronal signalling events. In this regard, a study using brain tissue has 20 recently shown that physiological concentrations of Pi can enhance the ATP-dependent binding of Ca++ to brain microsomes, resulting in a larger intracellular pool of Ca++ releasable by inositol triphosphate. Our recent work have demonstrated that >90% Pi transport in cortical neurons, 25 which displays similar kinetic parameters to those reported for cultured kidney proximal tubule epithelial cells and membrane vesicles, are sodium dependent and that this Na+dependent transport system is regulated through a Na+dependent Pi cotransporter. B. Ni, et al., Proceedings of 30 the National Academy of Sciences (USA), 91:5607-5611 (1994).

The present invention describes the cloning and characterization of a human brain Na^+ -dependent P_i cotransporter which is selectively expressed in discrete populations of neurons and glia. Fluorescent in situ hybridization (FISH) analysis demonstrates that this Na^+ -dependent P_i cotransporter is located in chromosome 19

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(19q13.3) which has been linked to susceptible gene(s) for late onset Alzheimer's disease. M. Mullan and F. Crawford, Trends in Neurological Sciences, 16, 398-403 (1993). The characterization and treatment of physiological disorders is hereby furthered.

This invention provides an isolated amino acid compound useful as a human brain sodium-dependent inorganic phosphate cotransporter, said compound comprising the amino acid sequence

	Met 1	Glu	Phe	Arg	Gln 5	Glu	Glu	Phe	Arg	Lys 10	Leu	Ala	Gly	Arg	Ala 15	Leu
15	Gly	Lys	Leu	His 20	Arg	Leu	Leu	Glu	Lys 25	Arg	Gln	Glu	Gly	Ala 30	Glu	Thr
20	Leu	Glu	Leu 35	Ser	Ala	Asp	Gly	Arg 40	Pro	Val	Thr	Thr	Gln 45	Thr	Arg	Asp
	Pro	Pro 50	Val	Val	Asp	Суз	Thr 55	Суз	Phe	Gly	Leu	Pro 60	Arg	Arg	Tyr	Ile
25	Ile 65	Ala	Ile	Met	Ser	Gly 70	Leu	Gly	Phe	Cys	Ile 75	Ser	Phe	Gly	Ile	Arg 80
	Cys	Asn	Leu	Gly	Val 85	Ala	Ile	Val	Ser	Met 90	Val	Asn	Asn	Ser	Thr 95	Thr
30	His	Arg	Gly	Gly 100	His	Val	Val	Val	Gln 105	Lys	Ala	Gln	Phe	Ser 110	Trp	Asp
35	Pro	Glu	Thr 115	Val	Gly	Leu	Ile	His 120	Gly	Ser	Phe	Phe	Trp 125	Gly	Tyr	Ile
	Val	Thr 130	Gln	Ile	Pro	Gly	Gly 135	Phe	Ile	Cys	Gln	Lys 140	Phe	Ala	Ala	Asn
40	Arg 145	Val	Phe	Gly	Phe	Ala 150	Ile	Val	Ala	Thr	Ser 155	Thr	Leu	Asn	Met	Leu 160
	Ile	Pro	Ser	Ala	Ala 165	Arg	Val	His	Tyr	Gly 170	Суѕ	Val	Ile	Phe	Val 175	Arg
45	Ile	Leu	Gln	Gly 180	Leu	Val	Glu	Gly	Val 185	Thr	Tyr	Pro	Ala	Cys 190	His	Gly
50	Ile	Trp	Ser 195	Lys	Trp	Ala	Pro	Pro 200	Leu	Glu	Arg	Ser	Arg 205	Leu	Ala	Thr
50																

	Thr	Ala 210	Phe	Cys	Gly	Ser	Tyr 215	Ala	Gly	Ala	Val	Va1 220	Ala	Met	Pro	Leu
5	Ala 225	Gly	Val	Leu	Val	Gln 230	Tyr	Ser	Gly	Trp	Ser 235	Ser	Val	Phe	Tyr	Val 240
	Tyr	Gly	Ser	Phe	Gly 245	Ile	Phe	Trp	Tyr	Leu 250	Phe	Trp	Leu	Leu	Val 255	Ser
10	Tyr	Glu	Ser	Pro 260	Ala	Leu	His	Pro	Ser 265	Ile	Ser	Glu	Glu	Glu 270	Arg	Lys
15	Tyr	Ile	Glu 275	Asp	Ala	Ile	Gly	Glu 280	Ser	Ala	Lys	Leu	Met 285	Asn	Pro	Leu
13	Thr	Lys 290	Phe	Ser	Thr	Pro	Trp 295	Arg	Arg	Phe	Phe	Thr 300	Ser	Met	Pro	Val
20	Tyr 305	Ala	Ile	Ile	Val	Ala 310	Asn	Phe	Cys	Arg	Ser 315	Trp	Thr	Phe	Tyr	Leu 320
	Leu	Leu	Ile	Ser	Gln 325	Pro	Asp	Tyr	Phe	Glu 330	Glu	Val	Phe	Gly	Phe 335	Glu
25	Ile	Ser	Lys	Val 340	Gly	Leu	Val	Ser	Ala 345	Leu	Pro	His	Leu	Val 350	Met	Thr
30	Ile	Ile	Val 355	Pro	Ile	Gly	Gly	Gln 360	Ile	Ala	Asp	Phe	Leu 365	Arg	Ser	Arg
30	Arg	Ile 370	Met	Ser	Thr	Thr	Asn 375	Val	Arg	Lys	Leu	Met 380	Asn	Суз	Gly	Gly
35	Phe 385	Gly	Met	Glu	Ala	Thr 390	Leu	Leu	Leu	Val	Val 395	Gly	Tyr	Ser	His	Ser 400
	Lys	Gly	Val	Ala	Ile 405	Ser	Phe	Leu	Val	Leu 410	Ala	Val	Gly	Phe	Ser 415	Gly
40	Phe	Ala	Ile	Ser 420	Gly	Phe	Asn	Val	Asn 425	His	Leu	Asp	Ile	Ala 430	Pro	Arg
45	Tyr	Ala	Ser 435	Ile	Leu	Met	Gly	Ile 440	Ser	Asn	Gly	Val	Gly 445	Thr	Leu	Ser
40	Gly	Met 450	Val	Cys	Pro	Ile	Ile 455	Val	Gly	Ala	Met	Thr 460	Lys	His	Lys	Thr
50	Arg 465	Glu	Glu	Trp	Gln	Tyr 470	Val	Phe	Leu	Ile	Ala 475	Ser	Leu	Val	His	Tyr 480
	Gly	Gly	Val	Ile	Phe 485	Tyr	Gly	Val	Phe	Ala 490	Ser	Gly	Glu	Lys	Gln 495	Pro
55	Trp	Ala	Glu	Pro	Glu	Glu	Met	Ser	Glu	Glu	Lys	Cys	Gly	Phe	Val	Gly

- 5 -

		500	505	510
-	His Asp Gln 515		Asp Asp Ser Glu Met Glu 520 525	Asp Glu Ala
5	Glu Pro Pro 530	Gly Ala Pro Pro 2 535	Ala Pro Pro Pro Ser Tyr 540	Gly Ala Thr
10	His Ser Thr 545	Phe Gln Pro Pro 1 550	Arg Pro Pro Pro Pro Val 555	Arg Asp Tyr 560
	hereinafter desi	ignated as SEQ	ID NO:2.	
	The i	nvention also	provides an isolated	nucleic
	acid compound th	nat comprises a	nucleic acid seque	nce which
15	encodes for the	amino acid com	mpounds provided. Pa	articularly
	_		solated nucleic acid	compound
	having the seque	ence		
20	CGATAAGCTT GATATO	CGAAT TCCGGACTCT	TGCTCGGGCG CCTTAACCCG G	CGTTCGGTT 60
20	CATCCCGCAG CGCCAG	STTCT GCTTACCAAA	AGTGGCCCAC TAGGCACTCG C	ATTCCACGC 120
	CCGCCTCCAC GCCAGC	GAGC CGGGCTTCTT	ACCCATTTAA AGTTTGAGAA T	AGGTTGAGA 180
25	TCGTTTCGGC CCCAAG	SACCT CTAATCATTC	GCTTTACCGG ATAAAACTGC G	TGGCGGGG 240
	TGCGTCGGGT CTGCG	AGAGC GCCAGCTATC	CTGAGGGAAA CTTCGGAGGG A	ACCAGCTAC 300
20	TAGATGGTTC GATTAG	STOTT TOGCCCCTAT	ACCCAGGTCG GACGACCGAT T	TGCACGTCA 360
30	GGACCGCTAC GGACCT	CCAC CAGAGTTTCC	TCTGGCTTCG CCCTGCCCAG G	CGATCGCCG 420
	GGGGGACCC GCGGGG	TGAC CGGCGGCAGG	AGCCGCCACC ATG GAG TTC	
35	,		Met Glu Phe . 1	Arg Gln 5
	GAG GAG TTT CGG A	AG CTA GCG GGT C	GT GCT CTC GGG AAG CTG	CAC CGC 523
	Glu Glu Phe Arg I	ys Leu Ala Gly A 10	rg Ala Leu Gly Lys Leu 1 15	His Arg 20
40				
			CG GAG ACG CTG GAG CTG . la Glu Thr Leu Glu Leu :	
	25		35	
45			CC CGG GAC CCG CCG GTG	
	Asp Gly Arg Pro V	al Thr Thr Gln T	hr Arg Asp Pro Pro Val ' 50	/al Asp
			GC TAC ATT ATC GCC ATC	
50	Cys Thr Cys Phe G 55	Cly Leu Pro Arg A: 60	rg Tyr Ile Ile Ala Ile I 65	1et Ser

5			GGC Gly		-						-				715
J			GTC Val												763
10			GTG Val												811
15	-		CAC His 120												859
20			TTT Phe												907
25			GTG Val												955
25			CAC His												1003
30			GGG Gly												1051
35			CCC Pro 200												1099 .
40			GCT Ala	Gly	Ala	Val	Val	Ala	Met	Pro		Alá	Gly		1147
45		Tyr	TCA Ser												1195
33			TGG Trp												1243
50			CCC Pro												1291
55			GAG Glu												1339

CCC TGG CGG CGC TTC TTC ACG TCT ATG CCA GTC TAT GCC ATC ATC GTG Pro Trp Arg Arg Phe Phe Thr Ser Met Pro Val Tyr Ala Ile Ile Val GCC AAC TTC TGC CGC AGC TGG ACG TTC TAC CTG CTG CTC ATC TCC CAG Ala Asn Phe Cys Arg Ser Trp Thr Phe Tyr Leu Leu Leu Ile Ser Gln CCC GAC TAC TTC GAA GAA GTG TTC GGC TTC GAG ATC AGC AAG GTA GGC Pro Asp Tyr Phe Glu Glu Val Phe Gly Phe Glu Ile Ser Lys Val Gly CTG GTG TCC GCG CTG CCC CAC CTG GTC ATG ACC ATC ATC GTG CCC ATC Leu Val Ser Ala Leu Pro His Leu Val Met Thr Ile Ile Val Pro Ile GGC GGC CAG ATC GCG GAC TTC CTG CGG AGC CGC CGC ATC ATG TCC ACC Gly Gly Gln Ile Ala Asp Phe Leu Arg Ser Arg Arg Ile Met Ser Thr ACC AAC GTG CGC AAG TTG ATG AAC TGC GGA GGC TTC GGC ATG GAA GCC Thr Asn Val Arg Lys Leu Met Asn Cys Gly Gly Phe Gly Met Glu Ala ACG CTG CTG GTG GTC GGC TAC TCG CAC TCC AAG GGC GTG GCC ATC Thr Leu Leu Leu Val Val Gly Tyr Ser His Ser Lys Gly Val Ala Ile TCC TTC CTG GTC CTA GCC GTG GGC TTC AGC GGC TTC GCC ATC TCT GGG Ser Phe Leu Val Leu Ala Val Gly Phe Ser Gly Phe Ala Ile Ser Gly TTC AAC GTG AAC CAC CTG GAC ATA GCC CCG CGC TAC GCC AGC ATC CTC Phe Asn Val Asn His Leu Asp Ile Ala Pro Arg Tyr Ala Ser Ile Leu ATG GGC ATC TCC AAC GGC GTG GGC ACA CTG TCG GGC ATG GTG TGC CCC Met Gly Ile Ser Asn Gly Val Gly Thr Leu Ser Gly Met Val Cys Pro ATC ATC GTG GGG GCC ATG ACT AAG CAC AAG ACT CGG GAG GAG TGG CAG Ile Ile Val Gly Ala Met Thr Lys His Lys Thr Arg Glu Glu Trp Gln TAC GTG TTC CTA ATT GCC TCC CTG GTG CAC TAT GGA GGT GTC ATC TTC Tyr Val Phe Leu Ile Ala Ser Leu Val His Tyr Gly Gly Val Ile Phe TAC GGG GTC TTT GCT TCT GGA GAG AAG CAG CCG TGG GCA GAG CCT GAG Tyr Gly Val Phe Ala Ser Gly Glu Lys Gln Pro Trp Ala Glu Pro Glu GAG ATG AGC GAG GAG AAG TGT GGC TTC GTT GGC CAT GAC CAG CTG GCT

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	Glu Met Ser Glu Glu Lys Cys Gly Phe Val Gly His Asp Gln Leu Ala 505 510 515	•
5	GGC AGT GAC GAC AGC GAA ATG GAG GAT GAG GCT GAG CCC CCG GGG GCA Gly Ser Asp Asp Ser Glu Met Glu Asp Glu Ala Glu Pro Pro Gly Ala 520 525 530	2059
10	CCC CCT GCA CCC CCG CCC TCC TAT GGG GCC ACA CAC AGC ACA TTT CAG Pro Pro Ala Pro Pro Pro Ser Tyr Gly Ala Thr His Ser Thr Phe Gln 535 540 545	2107
15	CCC CCC AGG CCC CCA CCC CCT GTC CGG GAC TAC TGA CCATGTGCCT Pro Pro Arg Pro Pro Pro Val Arg Asp Tyr * 550 555 560	2153
15	CCCACTGAAT GGCAGTTTCC AGGACCTCCA TTCCACTCAT CTCTGGCCTG AGTGACAGTG	2213
	TCAAGGAACC CTGCTCCTCT CTGTCCTGCC TCAGGCCTAA GAAGCACTCT CCCTTGTTCC	2273
20	CAGTGCTGTC AAATCCTCTT TCCTTCCCAA TTGCCTCTCA GGGGTAGTGA AGCTGCAGAC	2333
	TGACAGTTTC AAGGATACCC AAATTCCCCT AAAGGTTCCC TCTCCACCCG TTCTGCCTCA	2393
25	GTGGTTTCAA ATCTCTCCTT TCAGGGCTTT ATTTGAATGG ACAGTTCGAC CTCTTACTCT	2453
	CTCTTGTGGT TTTGAGGCAC CCACACCCC CGCTTTCCTT TATCTCCAGG GACTCTCAGG	2513
	CTAACCTTTG AGATCACTCA GCTCCCATCT CCTTTCAGAA AAATTCAAGG TCCTCCTCTA	2573
30	GAAGTTTCAA ATCTCTCCCA ACTCTGTTCT GCATCTTCCA GATTGGTTTA ACCAATTACT	2633
	CGTCCCCGCC ATTCCAGGGA TTGATTCTCA CCAGCGTTTC TGATGGAAAA TGGCGGGAAT	2693
	TCCTGCAGCC CGGGGGATCC ACT	2716

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which is hereinafter designated as SEQ ID NO:1.

This invention also provides recombinant nucleic acid vectors comprising nucleic acids encoding SEQ ID NO:2. This invention also encompasses recombinant DNA vectors which comprise the isolated DNA sequence which is SEQ ID NO:1.

The present invention also provides assays for determining the efficacy and adverse reaction profile of agents useful in the treatment or prevention of disorders associated with an inappropriate stimulation of a human brain Na*-dependent inorganic phosphate cotransporter.

The terms and abbreviations used in this document have their normal meanings unless otherwise designated. For

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example "'C" refers to degrees Celsius; "N" refers to normal or normality; "mmol" refers to millimole or millimoles; "g" refers to gram or grams; "ml" means milliliter or milliliters; "M" refers to molar or molarity; "µg" refers to microgram or micrograms; and "µl" refers to microliter or microliters.

All nucleic acid sequences, unless otherwise designated, are written in the direction from the 5' end to the 3' end, frequently referred to as "5' to 3'".

All amino acid or protein sequences, unless otherwise designated, are written commencing with the amino terminus ("N-terminus") and concluding with the carboxy terminus ("C-terminus").

"Base pair" or "bp" as used herein refers to DNA or 15 RNA. The abbreviations A,C,G, and T correspond to the 5'monophosphate forms of the deoxyribonucleosides (deoxy) adenine, (deoxy) cytidine, (deoxy) guanine, and (deoxy) thymine, respectively, when they occur in DNA molecules. The abbreviations U,C,G, and T correspond to the 20 5'-monophosphate forms of the ribonucleosides uracil, cytidine, guanine, and thymine, respectively when they occur in RNA molecules. In double stranded DNA, base pair may refer to a partnership of A with T or C with G. DNA/RNA, heteroduplex base pair may refer to a partnership of 25 A with U or C with G. (See the definition of "complementary", infra.)

The terms "digestion" or "restriction" of DNA refers to the catalytic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA ("sequence-specific endonucleases"). The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors, and other requirements were used as would be known to one of ordinary skill in the art. Appropriate buffers and substrate amounts for particular restriction enzymes are specified by the manufacturer or can be readily found in the literature.

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"Ligation" refers to the process of forming phosphodiester bonds between two double stranded nucleic acid fragments. Unless otherwise provided, ligation may be accomplished using known buffers and conditions with a DNA ligase, such as T4 DNA ligase.

The term "plasmid" refers to an extrachromosomal (usually) self-replicating genetic element. Plasmids are generally designated by a lower case "p" preceded and/or followed by letters and/or numbers. The starting plasmids herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accordance with published procedures. In addition, equivalent plasmids to those described are known in the art and will be apparent to the ordinarily skilled artisan.

The term "reading frame" means the nucleotide sequence from which translation occurs "read" in triplets by the translational apparatus of transfer RNA (tRNA) and ribosomes and associated factors, each triplet corresponding to a particular amino acid. A base pair insertion or deletion (termed a frameshift mutation) may result in two different proteins being coded for by the same DNA segment. To insure against this, the triplet codons corresponding to the desired polypeptide must be aligned in multiples of three from the initiation codon, i.e. the correct "reading frame" being maintained.

"Recombinant DNA cloning vector" as used herein refers to any autonomously replicating agent, including, but not limited to, plasmids and phages, comprising a DNA molecule to which one or more additional DNA segments can or have been added.

The term "recombinant DNA expression vector" as used herein refers to any recombinant DNA cloning vector in which a promoter to control transcription of the inserted DNA has been incorporated.

The term "expression vector system" as used herein refers to a recombinant DNA expression vector in combination

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with one or more trans-acting factors that specifically influence transcription, stability, or replication of the recombinant DNA expression vector. The trans-acting factor may be expressed from a co-transfected plasmid, virus, or other extrachromosomal element, or may be expressed from a gene integrated within the chromosome.

"Transcription" as used herein refers to the process whereby information contained in a nucleotide sequence of DNA is transferred to a complementary RNA sequence.

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The term "transfection" as used herein refers to the taking up of an expression vector by a host cell whether or not any coding sequences are in fact expressed. Numerous methods of transfection are known to the ordinarily skilled artisan, for example, calcium phosphate co-precipitation, and electroporation. Successful transfection is generally recognized when any indication of the operation of this vector occurs within the host cell.

The term "transformation" as used herein means the
introduction of DNA into an organism so that the DNA is
replicable, either as an extrachromosomal element or by
chromosomal integration. Methods of transforming bacterial
and eukaryotic hosts are well known in the art, many of which
methods, such as nuclear injection, protoplast fusion or by
calcium treatment using calcium chloride are summarized in J.
Sambrook, et al., MOLECULAR CLONING: A LABORATORY MANUAL, (1989).

The term "translation" as used herein refers to the process whereby the genetic information of messenger RNA is used to specify and direct the synthesis of a polypeptide chain.

The term "vector" as used herein refers to a nucleic acid compound used for the transformation of cells in gene manipulation bearing polynucleotide sequences corresponding to appropriate protein molecules which when combined with appropriate control sequences confer specific properties on the host cell to be transformed. Plasmids, viruses, and bacteriophage are suitable vectors. Artificial

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vectors are constructed by cutting and joining DNA molecules from different sources using restriction enzymes and ligases. The term "vector" as used herein includes Recombinant DNA cloning vectors and Recombinant DNA expression vectors.

The terms "complementary" or "complementarity" as used herein refers to pair of bases, purines and pyrimidines, that associate through hydrogen bonding in double stranded nucleic acid. The following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil.

The term "hybridization" as used herein refers to a process in which a strand of nucleic acid joins with a complementary strand through base pairing. The conditions employed in the hybridization of two non-identical, but very similar, complementary nucleic acids varies with the degree of complementarity of the two strands and the length of the strands. Such techniques and conditions are well known to practitioners in this field.

"Isolated amino acid sequence" refers to any amino acid sequence, however constructed or synthesized, which is locationally distinct from the naturally occurring sequence.

"Isolated DNA compound" refers to any DNA sequence, however constructed or synthesized, which is locationally distinct from its natural location in genomic DNA.

"Isolated nucleic acid compound" refers to any RNA or DNA sequence, however constructed or synthesized, which is locationally distinct from its natural location.

A "primer" is a nucleic acid fragment which functions as an initiating substrate for enzymatic or synthetic elongation.

The term "promoter" refers to a DNA sequence which directs transcription of DNA to RNA.

A "probe" as used herein is a nucleic acid compound or a fragment thereof which hybridizes with a nucleic acid compound which encodes either the entire sequence SEQ ID NO:2, a sequence complementary to SEQ ID NO:2, or a part thereof.

The term "stringency" refers to a set of hybridization conditions which may be varied in order to vary the degree of nucleic acid affinity for other nucleic acid. (See the definition of "hybridization", supra.)

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The term "antigenically distinct" as used herein refers to a situation in which antibodies raised against an epitope of the proteins of the present invention, or a fragment thereof, may be used to differentiate between the proteins of the present invention and other brain Na⁺⁻ dependent inorganic phosphate cotransporter subtypes. This term may also be employed in the sense that such antibodies may be used to differentiate between the human hBNPI protein protein and analogous proteins derived from other species.

The term "PCR" as used herein refers to the widely-15 known polymerase chain reaction employing a thermally-stable polymerase.

Skilled artisans will recognize that the proteins of the present invention can be synthesized by a number of different methods. All of the amino acid compounds of the invention can be made by chemical methods well known in the art, including solid phase peptide synthesis, or recombinant methods. Both methods are described in U.S. Patent 4,617,149, the entirety of which is herein incorporated by reference.

The principles of solid phase chemical synthesis of polypeptides are well known in the art and may be found in general texts in the area. See, e.g., H. Dugas and C. Penney, BIOORGANIC CHEMISTRY, (1981) at pages 54-92. For examples, peptides may be synthesized by solid-phase methodology utilizing an Applied Biosystems 430A peptide synthesizer (commercially available from Applied Biosystems, Foster City California) and synthesis cycles supplied by Applied Biosystems. Protected amino acids, such as t-butoxycarbonyl-protected amino acids, and other reagents are commercially available from many chemical supply houses.

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Sequential t-butoxycarbonyl chemistry using double couple protocols are applied to the starting p-methyl benzhydryl amine resins for the production of C-terminal carboxamides. For the production of C-terminal acids, the corresponding pyridine-2-aldoxime methiodide resin is used. Asparagine, glutamine, and arginine are coupled using preformed hydroxy benzotriazole esters. The following side chain protection may be used:

Arg, Tosyl

10 Asp, cyclohexyl

Glu, cyclohexyl

Ser, Benzyl

Thr, Benzyl

Tyr, 4-bromo carbobenzoxy

Removal of the t-butoxycarbonyl moiety
(deprotection) may be accomplished with trifluoroacetic acid
(TFA) in methylene chloride. Following completion of the
synthesis the peptides may be deprotected and cleaved from
the resin with anhydrous hydrogen fluoride containing 10%
meta-cresol. Cleavage of the side chain protecting group(s)
and of the peptide from the resin is carried out at zero
degrees centigrade or below, preferably -20°C for thirty
minutes followed by thirty minutes at 0°C.

After removal of the hydrogen fluoride, the peptide/resin is washed with ether, and the peptide extracted with glacial acetic acid and then lyophilized. Purification is accomplished by size-exclusion chromatography on a Sephadex G-10 (Pharmacia) column in 10% acetic acid.

The proteins of the present invention may also be produced by recombinant methods. Recombinant methods are preferred if a high yield is desired. A general method for the construction of any desired DNA sequence is provided in J. Brown, et al., Methods in Enzymology, 68:109 (1979). See also, J. Sambrook, et al., supra.

The basic steps in the recombinant production of desired proteins are:

a)	const	truct	cion	of	a	syn	thetic	or	semi-
synth	etic	DNA	enco	din	ıg	the	protei	n c	f
inter	est;								

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b) integrating said DNA into an expression vector in a manner suitable for the expression of the protein of interest, either alone or as a fusion protein;

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c) transforming an appropriate eukaryotic or prokaryotic host cell with said expression vector,

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d) culturing said transformed or transfected host cell in a manner to express the protein of interest; and

e) recovering and purifying the recombinantly produced protein of interest.

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In general, prokaryotes are used for cloning of DNA sequences in constructing the vectors of this invention. Prokaryotes may also be employed in the production of the protein of interest. For example, the <u>Escherichia coli K12</u> strain 294 (ATCC No. 31446) is particularly useful for the prokaryotic expression of foreign proteins. Other strains of <u>E. coli</u> which may be used (and their relevant genotypes) include the following.

30 Strain

Genotype

 $DH5\alpha$

F⁻ (ϕ 80dlacZ Δ M15), Δ (lacZYA-argF)U169 supE44, λ ⁻, hsdR17(r_K ⁻, m_K ⁺), recAl, endAl, gyrA96, thi-1, relAl

	нв101	supE44, hsdS20($r_B^ m_{B^-}$), recA13, ara-14, proA2 lacY1, galK2, rpsL20, xy1-5, mtl-1, mcrB, mrr
5	JM109	recA1, e14 ⁻ (mcrA), supE44, endA1, hsdR17(r_K^- , m_K^+), gyrA96, relA1, thi-1, Δ (lac-proAB), F'[traD36, proAB+ lacIq,lacZ Δ M15]
10	RR1	supE44, hsdS20($r_B^ m_B^-$), ara-14 proA2, lacY1, galK2, rpsL20, xyl-5, mtl-5
	χ1776	F^- , ton, A53, dapD8, minA1, supE42 (glnV42), Δ (gal-uvrB)40, minB2, rfb-
15		2, gyrA25, thyA142, oms-2, metC65, oms-1, Δ (bioH-asd)29, cycB2, cycA1, hsdR2, λ -
20	294	endA, thi ⁻ , hsr ⁻ , hsm _k ⁺ (U.S. Patent $4,366,246$)
	LE392	F-, hsdR514 (r-m-), supE44, supF58, lacY1, or Δ lac(I-Y)6, galK2, glaT22, metB1, trpR55, λ -
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These strains are all commercially available from suppliers such as: Bethesda Research Laboratories, Gaithersburg, Maryland 20877 and Stratagene Cloning Systems, La Jolla, California 92037; or are readily available to the poblic from sources such as the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland, 10852-1776.

Except where otherwise noted, these bacterial strains can be used interchangeably. The genotypes listed 35 are illustrative of many of the desired characteristics for choosing a bacterial host and are not meant to limit the

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invention in any way. The genotype designations are in accordance with standard nomenclature. See, for example, J. Sambrook, et al., supra. A preferred strain of E. coli employed in the cloning and expression of the genes of this invention is RV308, which is available from the ATCC under accession number ATCC 31608, and is described in United States Patent 4,551,433, issued November 5, 1985.

In addition to the strains of <u>E. coli</u> discussed <u>supra</u>, bacilli such as <u>Bacillus subtilis</u>, other enterobacteriaceae such as <u>Salmonella typhimurium</u> or <u>Serratia marcescans</u>, and various <u>Pseudomonas</u> species may be used. In addition to these gram-negative bacteria, other bacteria, especially <u>Streptomyces</u>, spp., may be employed in the prokaryotic cloning and expression of the proteins of this invention.

Promoters suitable for use with prokaryotic hosts include the β -lactamase [vector pGX2907 (ATCC 39344) contains the replicon and β -lactamase gene] and lactose promoter systems [Chang et al., Nature (London), 275:615 (1978); and 20 Goeddel et al., Nature (London), 281:544 (1979)], alkaline phosphatase, the tryptophan (trp) promoter system [vector pATH1 (ATCC 37695) is designed to facilitate expression of an open reading frame as a trpE fusion protein under control of the trp promoter] and hybrid promoters such as the tac 25 promoter (isolatable from plasmid pDR540 ATCC-37282). However, other functional bacterial promoters, whose nucleotide sequences are generally known, enable one of skill in the art to ligate them to DNA encoding the proteins of the instant invention using linkers or adapters to supply any 30 required restriction sites. Promoters for use in bacterial systems also will contain a Shine-Dalgarno sequence operably linked to the DNA encoding the desired polypeptides. examples are illustrative rather than limiting.

The proteins of this invention may be synthesized either by direct expression or as a fusion protein comprising the protein of interest as a translational fusion with another protein or peptide which may be removable by

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enzymatic or chemical cleavage. It is often observed in the production of certain peptides in recombinant systems that expression as a fusion protein prolongs the lifespan, increases the yield of the desired peptide, or provides a convenient means of purifying the protein of interest. A variety of peptidases (e.g. trypsin) which cleave a polypeptide at specific sites or digest the peptides from the amino or carboxy termini (e.g. diaminopeptidase) of the peptide chain are known. Furthermore, particular chemicals (e.g. cyanogen bromide) will cleave a polypeptide chain at specific sites. The skilled artisan will appreciate the modifications necessary to the amino acid sequence (and synthetic or semi-synthetic coding sequence if recombinant means are employed) to incorporate site-specific internal cleavage sites. See e.g., P. Carter, "Site Specific Proteolysis of Fusion Proteins", Chapter 13 in PROTEIN PURIFICATION: FROM MOLECULAR MECHANISMS TO LARGE SCALE PROCESSES, American Chemical Society, Washington, D.C. (1990).

In addition to cloning and expressing the genes of interest in the prokaryotic systems discussed above, the proteins of the present invention may also be produced in eukaryotic systems. The present invention is not limited to use in a particular eukaryotic host cell. A variety of eukaryotic host cells are available from depositories such as the American Type Culture Collection (ATCC) and are suitable for use with the vectors of the present invention. The choice of a particular host cell depends to some extent on the particular expression vector used to drive expression of the nucleic acids of the present invention. Exemplary host cells suitable for use in the present invention are listed in Table I.

Table I

Host Cell	Origin	Source
HepG-2	Human Liver Hepatoblastoma	АТСС НВ 8065

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CV-1	African Green Monkey Kidney	ATCC CCL 70
LLC-MK ₂	Rhesus Monkey Kidney	ATCC CCL 7.1
3T 3	Mouse Embryo Fibroblasts	ATCC CCL 92
СНО-К1	Chinese Hamster Ovary	ATCC CCL 61
HeLa	Human Cervix Epitheloid	ATCC CCL 2
RPMI8226	Human Myeloma	ATCC CCL 155
H4IIEC3	Rat Hepatoma	ATCC CCL 1600
C127I	Mouse Fibroblast	ATCC CCL 1616
HS-Sultan	Human Plasma Cell Plasmocytoma	ATCC CCL 1484
внк-21	Baby Hamster Kidney	ATCC CCL 10

An especially preferred cell line employed in this invention is the widely available cell line AV12-664 (hereinafter "AV12"). This cell line is available from the American Type Culture Collection under the accession number ATCC CRL 9595. The AV12 cell line was constructed by injecting a Syrian hamster in the scruff of the neck with human adenovirus 12 and isolating cells from the resulting tumor.

A wide variety of vectors, some of which are discussed below, exists for the transformation of such mammalian host cells, but the specific vectors described herein are in no way intended to limit the scope of the present invention.

The pSV2-type vectors comprise segments of the simian virus 40 (SV40) genome that constitute a defined eukaryotic transcription unit-promoter, intervening sequence, and polyadenylation site. In the absence of the SV40 T antigen, the plasmid pSV2-type vectors transform mammalian and other eukaryotic host cells by integrating into the host cell chromosomal DNA. A large number of plasmid pSV2-type vectors have been constructed, such as plasmid pSV2-gpt, pSV2-neo, pSV2-dhfr, pSV2-hyg, and pSV2- β -globin, in which the SV40 promoter drives transcription of an inserted gene. These vectors are suitable for use with the coding sequences of the present invention and are widely available from

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sources such as the ATCC or the Northern Regional Research Laboratory (NRRL), 1815 N. University Street, Peoria, Illinois, 61604.

The plasmid pSV2-dhfr (ATCC 37146) comprises a murine dihydrofolate reductase (dhfr) gene under the control of the SV40 early promoter. Under the appropriate conditions, the dhfr gene is known to be amplified, or copied, in the host chromosome. This amplification can result in the amplification of closely-associated DNA sequences and can, therefore, be used to increase production of a protein of interest. See. e.g., J. Schimke, Cell, 35:705-713 (1984).

Plasmids constructed for expression of the proteins of the present invention in mammalian and other eukaryotic host cells can utilize a wide variety of promoters. present invention is in no way limited to the use of the particular promoters exemplified herein. Promoters such as the SV40 late promoter, promoters from eukaryotic genes, such as, for example, the estrogen-inducible chicken ovalbumin gene, the interferon genes, the gluco-corticoid-inducible tyrosine aminotransferase gene, and the thymidine kinase gene, and the major early and late adenovirus genes can be readily isolated and modified to express the genes of the present invention. Eukaryotic promoters can also be used in tandem to drive expression of a coding sequence of this invention. Furthermore, a large number of retroviruses are known that infect a wide range of eukaryotic host cells. long terminal repeats in the retroviral DNA frequently encode functional promoters and, therefore, may be used to drive expression of the nucleic acids of the present invention.

Plasmid pRSVcat (ATCC 37152) comprises portions of a long terminal repeat of the Rous Sarcoma virus, a virus known to infect chickens and other host cells. This long terminal repeat contains a promoter which is suitable for use in the vectors of this invention. H. Gorman, et al., Proceedings of the National Academy of Sciences (USA), 79:6777 (1982). The plasmid pMSVi (NRRL B-15929) comprises

the long terminal repeats of the Murine Sarcoma virus, a virus known to infect mouse and other host cells. The mouse metallothionein promoter has also been well characterized for use in eukaryotic host cells and is suitable for use in the expression of the nucleic acids of the present invention. The mouse metallothionein promoter is present in the plasmid pdBPV-MMTneo (ATCC 37224) which can serve as the starting material of other plasmids of the present invention.

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An especially preferred expression vector system employs one of a series of vectors containing the BK enhancer, an enhancer derived from the BK virus, a human papovavirus. The most preferred such vector systems are those which employ not only the BK enhancer but also the adenovirus-2-early region 1A (E1A) gene product. The E1A gene product (actually, the E1A gene produces two products, which are collectively referred to herein as "the E1A gene product") is an immediate-early gene product of adenovirus, a large DNA virus.

A most preferred expression vector employed in the 20 present invention is the phd series of vectors which comprise a BK enhancer in tandem with the adenovirus late promoter to drive expression of useful products in eukaryotic host cells. The construction and method of using the phd plasmid, as well as related plasmids, are described in U.S. Patents 5,242,688, 25 issued September 7, 1993, and 4,992,373, issued February 12, 1991, as well as co-pending United States patent application 07/368,700, all of which are herein incorporated by reference. Escherichia coli K12 GM48 cells harboring the plasmid phd are available as part of the permanent stock collection of the Northern Regional Research Laboratory under 30 accession number NRRL B-18525. The plasmid may be isolated from this culture using standard techniques.

The plasmid phd contains a unique <u>Bcl</u>I site which may be utilized for the insertion of the gene encoding the protein of interest. The skilled artisan understands that linkers or adapters may be employed in cloning the gene of interest into this <u>Bcl</u>I site. A depiction of the plasmid phd

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is provided as Figure 2 of this document. The phd series of plasmids functions most efficiently when introduced into a host cell which produces the EIA gene product, cell lines such as AV12-664, 293 cells, and others, described <u>supra</u>.

Transformation of the mammalian cells can be performed by any of the known processes including, but not limited to, the protoplast fusion method, the calcium phosphate co-precipitation method, electroporation and the like. See, e.g., J. Sambrook, et al., supra, at 3:16.30-3:16.66.

Other routes of production are well known to skilled artisans. In addition to the plasmid discussed above, it is well known in the art that some viruses are also appropriate vectors. For example, the adenovirus, the adeno-associated virus, the vaccinia virus, the herpes virus, the baculovirus, and the rous sarcoma virus are useful. Such a method is described in U.S. Patent 4,775,624, herein incorporated by reference. Several alternate methods of expression are described in J. Sambrook, et al., supra, at 16.3-17.44.

In addition to prokaryotes and mammalian host cells, eukaryotic microbes such as yeast cultures may also be used. The imperfect fungus <u>Saccharomyces cerevisiae</u>, or common baker's yeast, is the most commonly used eukaryotic microorganism, although a number of other strains are commonly available. For expression in <u>Saccharomyces</u> sp., the plasmid YRp7 (ATCC-40053), for example, is commonly used. <u>See. e.g.</u>, L. Stinchcomb, <u>et al.</u>, <u>Nature</u>, 282:39 (1979); J. Kingsman <u>et al.</u>, <u>Gene</u>, 7:141 (1979); S. Tschemper <u>et al.</u>, <u>Gene</u>, 10:157 (1980). This plasmid already contains the <u>trp</u> gene which provides a selectable marker for a mutant strain of yeast lacking the ability to grow in tryptophan.

Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase

[found on plasmid pAP12BD (ATCC 53231) and described in U.S. Patent No. 4,935,350, issued June 19, 1990, herein incorporated by reference] or other glycolytic enzymes such

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as enolase [found on plasmid pAC1 (ATCC 39532)], glyceraldehyde-3-phosphate dehydrogenase [derived from plasmid pHcGAPC1 (ATCC 57090, 57091)], hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase, as well as the alcohol dehydrogenase and pyruvate decarboxylase genes of Zymomonas mobilis (United States Patent No. 5,000,000 issued March 19, 1991, herein incorporated by reference).

Other yeast promoters, which are inducible promoters, having the additional advantage of their transcription being controllable by varying growth conditions, are the promoter regions for alcohol 15 dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein [contained on plasmid vector pCL28XhoLHBPV (ATCC 39475) and described in United States Patent No. 4,840,896, herein incorporated by reference], glyceraldehyde 20 3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose [e.g. GAL1 found on plasmid pRY121 (ATCC 37658)] utilization. Suitable vectors and promoters for use in yeast expression are further described in R. Hitzeman et al., European Patent Publication No. 73,657A. Yeast enhancers such as the UAS Gal from <u>Saccharomyces</u> 25 cerevisiae (found in conjuction with the CYCl promoter on plasmid YEpsec--hIlbeta ATCC 67024), also are advantageously used with yeast promoters.

Practitioners of this invention realize that, in

addition to the above-mentioned expression systems, the
cloned cDNA may also be employed in the production of
transgenic animals in which a test mammal, usually a mouse,
in which expression or overexpression of the proteins of the
present invention can be assessed. The nucleic acids of the
present invention may also be employed in the construction of
"knockout" animals in which the expression of the native
cognate of the gene is suppressed.

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Skilled artisans also recognize that some alterations of SEQ ID NO:2 will fail to change the function of the amino acid compound. For instance, some hydrophobic amino acids may be exchanged for other hydrophobic amino acids. Those altered amino acid compounds which confer substantially the same function in substantially the same manner as the exemplified amino acid compound are also encompassed within the present invention. Typical such conservative substitutions attempt to preserve the: (a) secondary or tertiary structure of the polypeptide backbone; (b) the charge or hydrophobicity of the residue; or (c) the bulk of the side chain. Some examples of such conservative substitutions of amino acids, resulting in the production of proteins which are functional equivalents of the protein of SEQ ID NO:2 are shown in Table II, infra.

Table II

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_	Original Residue	Exemplary Substitutions
	Ala	Ser, Gly
20	Arg	Lys
	Asn	Gln, His
	Asp	Glu
	Cys	Ser
	Gln	Asn
25	Glu	Asp
	Gly	Pro, Ala
	His	Asn, Gln
	lle	Leu, Val
	Leu	Ile, Val
30	Lys	Arg, Gln, Glu
	Mel	Leu, Ile
	Phe	Met, Leu, Gyr
	Ser	Thr
	Thr	Ser
35	Trp	Tyr

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Tyr Trp, Phe
Val Ile, Leu

These substitutions may be introduced into the protein in a variety of ways, such as during the chemical synthesis or by chemical modification of an amino acid side chain after the protein has been prepared.

Alterations of the protein having a sequence which corresponds to the sequence of SEQ ID NO:2 may also be induced by alterations of the nucleic acid compounds which encodes these proteins. These mutations of the nucleic acid compound may be generated by either random mutagenesis techniques, such as those techniques employing chemical mutagens, or by site-specific mutagenesis employing oligonucleotides. Those nucleic acid compounds which confer substantially the same function in substantially the same manner as the exemplified nucleic acid compounds are also encompassed within the present invention.

Other embodiments of the present invention are nucleic acid compounds which comprise isolated nucleic acid sequences which encode SEQ ID NO:2. As skilled artisans will recognize, the amino acid compounds of the invention can be encoded by a multitude of different nucleic acid sequences because most of the amino acids are encoded by more than one nucleic acid triplet due to the degeneracy of the amino acid code. Because these alternative nucleic acid sequences would encode the same amino acid sequences, the present invention further comprises these alternate nucleic acid sequences.

The gene encoding the hBNPI protein molecule may be produced using synthetic methodology. This synthesis of nucleic acids is well known in the art. See. e.g., E.L. Brown, R. Belagaje, M.J. Ryan, and H.G. Khorana, Methods in Enzymology, 68:109-151 (1979). The DNA segments corresponding to the receptor gene are generated using conventional DNA synthesizing apparatus such as the Applied Biosystems Model 380A or 380B DNA synthesizers (commercially available from Applied Biosystems, Inc., 850 Lincoln Center

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Drive, Foster City, CA 94404) which employ phosphoramidite chemistry. In the alternative, the more traditional phosphotriester chemistry may be employed to synthesize the nucleic acids of this invention. [See, e.g., M.J. Gait, ed., OLIGONUCLEOTIDE SYNTHESIS, A PRACTICAL APPROACH, (1984).]

The synthetic human hBNPI protein gene may be designed to possess restriction endonuclease cleavage sites at either end of the transcript to facilitate isolation from and integration into expression and amplification plasmids. The choice of restriction sites are chosen so as to properly orient the coding sequence of the receptor with control sequences to achieve proper in-frame reading and expression of the hBNPI protein. A variety of other such cleavage sites may be incorporated depending on the particular plasmid constructs employed and may be generated by techniques well known in the art.

In an alternative methodology, the desired DNA sequences can be generated using the polymerase chain reaction as described in U.S. Patent No. 4,889,818, which is herein incorporated by reference.

In addition to the deoxyribonucleic acid of SEQ ID NO:1, this invention also provides ribonucleic acids (RNA) which comprise the RNA sequence

25	CGAUAAGCUU	GAUAUCGAAU	UCCGGACUCU	UGCUCGGGCG	CCUUAACCCG	GCGUUCGGUU	60
	CAUCCCGCAG	CGCCAGUUCU	GCUUACCAAA	AGUGGCCCAC	UAGGCACUCG	CAUUCCACGC	120
30	CCGCCUCCAC	GCCAGCGAGC	CGGCCUUCUU	ACCCAUUUAA	AGUUUGAGAA	UAGGUUGAGA	180
30	UCGUUUCGGC	CCCAAGACCU	CUAAUCAUUC	GCUUUACCGG	AUAAAACUGC	GUGGCGGGG	240
	UGCGUCGGGU	CUGCGAGAGC	GCCAGCUAUC	CUGAGGGAAA	CUUCGGAGGG	AACCAGCUAC	300
35	UAGAUGGUUC	GAUUAGUCUU	UCGCCCCUAU	ACCCAGGUCG	GACGACCGAU	UUGCACGUCA	360
	GGACCGCUAC	GGACCUCCAC	CAGAGUUUCC	UCUGGCUUCG	CCCUGCCCAG	GCGAUCGGCG	420
40	GGGGGACCC	GCGGGGUGAC	CGGCGGCAGG	AGCCGCCACC	AUGGAGUUCC	GCCAGGAGGA	480
40	GUUUCGGAAG	CUAGCGGGUC	GUGCUCUCGG	GAAGCUGCAC	cgccuucugg	AGAAGCGGCA	540
	GGAAGGCGCG	GAGACGCUGG	AGCUGAGUGC	GGAUGGGCGC	CCGGUGACCA	CGCAGACCCG	600

	GGACCCGCCG	GUGGUGGACU	GCACCUGCUU	CGGCCUCCCU	CGCCGCUACA	UUAUCGCCAU	660
5	CAUGAGUGGU	CUGGGCUUCU	GCAUCAGCUU	UGGCAUCCGC	UGCAACCUGG	GCGUGGCCAU	720
	CGUCUCCAUG	GUCAAUAACA	GCACGACCCA	ccccccccc	CACGUGGUGG	UGCAGAAAGC	780
	CCAGUUCAGC	UGGGAUCCAG	AGACUGUCGG	CCUCAUACAC	GCCUCCUUUU	UCUGGGGCUA	840
10	CAUUGUCACU	CAGAUUCCAG	GAGGAUUUAU	CUGUCAAAAA	UUUGCAGCCA	ACAGAGUUUU	900
	CGGCUUUGCU	AUUGUGGCAA	CAUCCACUCU	AAACAUGCUG	AUCCCCUCAG	cugcccgcgu	960
15	CCACUAUGGC	UGUGUCAUCU	UCGUGAGGAU	CCUGCAGGGG	UUGGUAGAGG	GGGUCACAUA	1020
	CCCCGCCUGC	CAUGGGAUCU	GGAGCAAAUG	GGCCCCACCC	UUAGAACGGA	GUCGCCUGGC	1080
	GACGACAGCC	UUUUGUGGUU	CCUAUGCUGG	GGCGGUGGUC	GCGAUGCCCC	UCGCCGGGGU	1140
20	CCUUGUGCAG	UACUCAGGAU	GGAGCUCUGU	UUUCUACGUC	UACGGCAGCU	UCGGGAUCUU	1200
	CUGGUACCUG	UUCUGGCUGC	UCGUCUCCUA	CGAGUCCCCC	GCGCUGCACC	CCAGCAUCUC	1260
25	GGAGGAGGAG	CGCAAGUACA	UCGAGGACGC	CAUCGGAGAG	AGCGCGAAAC	UCAUGAACCC	1320
	CCUCACGAAG	UUUAGCACUC	CCUGGCGGCG	CUUCUUCACG	UCUAUGCCAG	UCUAUGCCAU	1380
	CAUCGUGGCC	AACUUCUGCC	GCAGCUGGAC	GUUCUACCUG	CUGCUCAUCU	CCCAGCCCGA	1440
30	CUACUUCGAA	GAAGUGUUCG	GCUUCGAGAU	CAGCAAGGUA	GCCUGGUGU	CCGCGCUGCC	1500
	CCACCUGGUC	AUGACCAUCA	UCGUGCCCAU	CGGCGGCCAG	AUCGCGGACU	UCCUGCGGAG	1560
35	CCGCCGCAUC	AUGUCCACCA	CCAACGUGCG	CAAGUUGAUG	AACUGCGGAG	GCUUCGGCAU	1620
	GGAAGCCACG	CUGCUGUUGG	UGGUCGGCUA	CUCGCACUCC	AAGGCGUGG	CCAUCUCCUU	1680
	CCUGGUCCUA	GCCGUGGGCU	UCAGCGGCUU	CGCCAUCUCU	GGGUUCAACG	UGAACCACCU	1740
40	GGACAUAGCC	CCGCGCUACG	CCAGCAUCCU	CAUGGGCAUC	UCCAACGGCG	UGGGCACACU	1800
	GUCGGGCAUG	GUGUGCCCCA	UCAUCGUGGG	GGCCAUGACU	AAGCACAAGA	CUCGGGAGGA	1860
45	GUGGCAGUAC	GUGUUCCUAA	UUGCCUCCCU	GGUGCACUAU	GGAGGUGUCA	UCUUCUACGG	1920
	GCUCUUUGCU	UCUGGAGAGA	AGCAGCCGUG	GGCAGAGCCU	GAGGAGAUGA	GCGAGGAGAA	1980
	GUGUGGCUUC	GUUGGCCAUG	ACCAGCUGGC	UGGCAGUGAC	GACAGCGAAA	UGGAGGAUGA	2040
50	GGCUGAGCCC	CCGGGGGCAC	CCCCUGCACC	ccccccccc	UAUGGGGCCA	CACACAGCAC	2100
	AUUUCAGCCC	CCCAGGCCCC	CACCCCCUGU	CCGGGACUAC	UGACCAUGUG	CCUCCCACUG	2160
55	AAUGGCAGUU	UCCAGGACCU	CCAUUCCACU	CAUCUCUGGC	CUGAGUGACA	GUGUCAAGGA	2220

	ACCCUGCUCC	ucucuguccu	GCCUCAGGCC	UAAGAAGCAC	UCUCCCUUGU	UCCCAGUGCU	2280
5	GUCAAAUCCU	cuuuccuucc	CAAUUGCCUC	UCAGGGGUAG	UGAAGCUGCA	GACUGACAGU	2340
	UUCAAGGAUA	CCCAAAUUCC	CCUAAAGGUU	CCCUCUCCAC	ccguucugcc	UCAGUGGUUU	2400
	CAAAUCUCUC	CUUUCAGGGC	UUUAUUUGAA	UGGACAGUUC	GACCUCUUAC	UCUCUCUUGU	2460
10	GGUUUUGAGG	CACCCACACC	ccccccuuuc	CUUUAUCUCC	AGGGACUCUC	AGGCUAACCU	2520
	UUGAGAUCAC	UCAGCUCCCA	UCUCCUUUCA	GAAAAAUUCA	AGGUCCUCCU	CUAGAAGUUU	2580
	CAAAUCUCUC	CCAACUCUGU	UCUGCAUCUU	CCAGAUUGGU	UUAACCAAUU	ACUCGUCCCC	2640
15	GCCAUUCCAG	GGAUUGAUUC	UCACCAGCGU	UUCUGAUGGA	AAAUGGCGGG	AAUUCCUGCA	2700
	GCCCGGGGGA	UCCACU					2716

hereinafter referred to as SEQ ID NO:3, or the complementary ribonucleic acid, or a fragment of either SEQ ID NO:3 or the complement thereof. The ribonucleic acids of the present invention may be prepared using the polynucleotide synthetic methods discussed <u>supra</u> or they may be prepared enzymatically using RNA polymerases to transcribe a DNA template.

25 complement thereof.

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The most preferred systems for preparing the ribonucleic acids of the present invention employ the RNA polymerase from the bacteriophage T7 or the bacteriophage SP6. Both of these RNA polymerases are highly specific and require the insertion of bacteriophage-specific sequences at the 5' end of the message to be read. See, J. Sambrook, et al., supra, at 18.82-18.84.

This invention also provides nucleic acids, RNA or DNA, which are complementary to SEQ ID NO:1 or SEQ ID NO:3.

The present invention also provides probes and primers useful for molecular biology techniques. A compound which encodes for SEQ ID NO:1, SEQ ID NO:3 or a complementary sequence of SEQ ID NO:1 or SEQ ID NO:3, or a fragment thereof, and which is at least 18 base pairs in length, and which will selectively hybridize to human genomic DNA or messenger RNA encoding a human brain Na+-dependent inorganic

phosphate cotransporter, is provided. Preferably, the 18 or more base pair compound is DNA.

The term "selectively hybridize" as used herein may refer to either of two situations. In the first such embodiment of this invention, the nucleic acid compounds described supra hybridize to a human sodium-dependent inorganic phosphate cotransporter under more stringent hybridization conditions than these same nucleic acid compounds would hybridize to an analogous sodium-dependent inorganic phosphate cotransporter of another species, e.g. murine or primate. In the second such embodiment of this invention, these probes hybridize to the hBNPI protein of the present invention under more stringent hybridization conditions than other related compounds, including nucleic acid sequences encoding other ion cotransporters.

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These probes and primers can be prepared enzymatically as described <u>supra</u>. In a most preferred embodiment these probes and primers are synthesized using chemical means as described <u>supra</u>. Probes and primers of defined structure may also be purchased commercially.

This invention also encompasses recombinant DNA cloning vectors and expression vectors comprising the nucleic acids of the present invention. Many of the vectors encompassed within this invention are described above. The preferred nucleic acid vectors are those which are DNA.

The sequence of SEQ ID NO:1 was prepared as follows:

Molecular cloning of a human brain Na±-dependent inorganic phosphate cotransporter(hBNPI)

Using a cDNA encoding the rat brain Na⁺-dependent inorganic phosphate cotransporter (rBNPI) (Ni, et al., 1994), we screened, under low stringency conditions, a human cDNA library derived from hippocampus mRNAs. Twelve positive clones were isolated that strongly hybridized to the ³²P-labeled probe rBNPI. Restriction endonuclease analysis

and/or sequencing of these clones revealed two distinct sequences: those which are highly similar to the rBNPI (B. Ni, et al., 1994, supra) as well as the kidney Na+-dependent inorganic phosphate cotransporter (Na/Pi), found in 10 clones, and those found in 2 clones which were proved to be rearrangments between the human putative phosphate transporter and other cDNAs. Of the 10 clones (designed as hBNP) which exhibited a strong similarity to rBNPI, 4 clones contained the 2.7 kb message. Sequence analysis of hBNPI 10 predicts an open reading frame of 1683 bases, corresponding to a protein of 560 amino acids with an apparent molecular mass of 61,000 Da (61 kDa). The ATG initiation codon at position 1, which is preceded by an upstream, in-frame stop codon, matches the Kazak consensus initiation sequence for the initiation of translation.

Computer searching revealed that the protein encoded by the hBNPI shared significant sequence homology at the amino acid level with those of recently cloned rat rBNPI (98%), rabbit (31%) and human (31%) kidney phosphate 20 transporter, Na/P_i, as indicated by comparison analysis. highest degree of homology, which was found between rBNPI and hBNPI, suggested that hBNPI is the human homologue of the rat rBNPI. The segment of highest homology among the proteins is confined to a region that fits the proposed consensus Na+-binding domain for various Na+-dependent 25 transporter systems (Deguchi et al., 1990). Alignment of the predicted hBNPI protein sequence with the consensus sequence indicated that amino acids leucine (L), glycine (G) and arginine (R) residues match the proposed motif and that other (F and R) are conservatively changed. The predicted hBNPI 30 protein sequence also shares 41% and 32% amino acid identity with two proteins of unknown function from Caenorhabditis elegans, ZK512.6 and C38C10.2, respectively. J. Sulston et al., Nature (London), 356:37-41 (1992). A hydropathy plot of the deduced amino acid sequence of hBNPI suggests the 35 presence of at least 6 to 8 transmembrane regions. This

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number of membrane-spanning domains is a characteristic structural motif of transport proteins. Based on the convention that activity of neuronal P_i transport correlates with ATP synthesis and intracellular energy charge, we have modelled hBNPI protein secondary structure with 6 transmembrane domains, which is consistent with those of other energy-linked anion transporters. The putative two glycosylation sites and two protein kinase C phosphorylation sites and four putative calmodulin-dependent kinase II phosphorylation sites are well conserved

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The skilled artisan understands that the type of cloning vector or expression vector employed depends upon the availability of appropriate restriction sites, the type of host cell in which the vector is to be transfected or transformed, the purpose of the transfection or transformation (e.g., transient expression in an oocyte system, stable transformation as an extrachromosomal element, or integration into the host chromosome), the presence or absence of readily assayable markers (e.g., antibiotic resistance markers, metabolic markers, or the like), and the number of copies of the gene to be present in the cell.

The type of vector employed to carry the nucleic acids of the present invention may be RNA viruses, DNA viruses, lytic bacteriophages, lysogenic bacteriophages, stable bacteriophages, plasmids, viroids, and the like. The most preferred vectors of the present invention are those derived from plasmids.

When preparing an expression vector the skilled artisan understands that there are many variables to be considered. One such example is the use of a constitutive promoter, i.e. a promoter which is functional at all times, instead of a regulatable promoter which may be activated or inactivated by the artisan using heat, addition or removal of a nutrient, addition of an antibiotic, and the like. The practitioner also understands that the amount of nucleic acid or protein to be produced dictates, in part, the selection of the expression system. For experiments examining the amount

of the protein expressed on the cell membrane or for experiments examining the biological function of an expressed membrane protein, for example, it may be unwise to employ an expression system which produces too much of the protein.

The addition or subtraction of certain sequences, such as a signal sequence preceding the coding sequence, may be employed by the practitioner to influence localization of the resulting polypeptide. Such sequences added to or removed from the nucleic acid compounds of the present invention are encompassed within this invention.

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The starting plasmids employed to prepare the vectors of the present invention may be isolated from the appropriate <u>E. coli</u> containing these plasmids using standard procedures such as cesium chloride DNA isolation.

The plasmids of the present invention may be readily modified to construct expression vectors that produce hBNPI proteins in a variety of organisms, including, for example, <u>E. coli</u>, Sf9 (as host for baculovirus), <u>Spodoptera</u> and <u>Saccharomyces</u>. The current literature contains

20 techniques for constructing AV12 expression vectors and for transforming AV12 host cells. United States Patent No. 4,992,373, herein incorporated by reference, is one of many references describing these techniques.

One of the most widely employed techniques for 25 altering a nucleic acid sequence is by way of oligonucleotide-directed site-specific mutagenesis. B. Comack, Current Protocols in Molecular Biology, 8.01-8.5.9, (F. Ausubel, et al., eds. 1991). In this technique an oligonucleotide, whose sequence contains the mutation of 30 interest, is synthesized as described <u>supra</u>. oligonucleotide is then hybridized to a template containing the wild-type sequence. In a most preferred embodiment of this technique, the template is a single-stranded template. Particularly preferred are plasmids which contain regions such as the f1 intergenic region. This region allows the 35 generation of single-stranded templates when a helper phage is added to the culture harboring the "phagemid".

After the annealing of the oligonucleotide to the template, a DNA-dependent DNA polymerase is then used to synthesize the second strand from the oliognucleotide, complementary to the template DNA. The resulting product is a heteroduplex molecule containing a mismatch due to the mutation in the oligonucleotide. After DNA replication by the host cell a mixture of two types of plasmid are present, the wild-type and the newly constructed mutant. This technique permits the introduction of convenient restriction sites such that the coding sequence may be placed immediately adjacent to whichever transcriptional or translational regulatory elements are employed by the practitioner.

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The construction protocols utilized for <u>E. coli</u> can be followed to construct analogous vectors for other organisms, merely by substituting, if necessary, the appropriate regulatory elements using techniques well known to skilled artisans.

Host cells which harbor the nucleic acids provided by the present invention are also provided. A preferred host cell is an Xenopus sp. oocyte which has been injected with RNA or DNA compounds of the present invention. Most preferred oocytes of the present invention are those which harbor a sense mRNA of the present invention. Other preferred host cells include AV12 and <u>E. coli</u> cells which have been transfected and/or transformed with a vector which comprises a nucleic acid of the present invention.

The present invention also provides a method for constructing a recombinant host cell capable of expressing SEQ ID NO:2, said method comprising transforming a host cell with a recombinant DNA vector that comprises an isolated DNA sequence which encodes SEQ ID NO:2. The preferred host cell is AV12. The preferred vector for expression is one which comprises SEQ ID NO:1. Another preferred host cell for this method is E. coli. An especially preferred expression vector in E. coli is one which comprises SEQ ID NO:1. Transformed host cells may be cultured under conditions well known to

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skilled artisans such that SEQ ID NO:2 is expressed, thereby producing Yb in the recombinant host cell.

The ability of ions to bind to the hBNPI protein is essential in the development of a multitude of indications. In developing agents which act as antagonists or agonists of the hBNPI protein, it would be desirable, therefore, to determine those agents which bind the hBNPI protein.

Generally, such an assay includes a method for determining whether a substance is a functional ligand of the hBNPI protein, said method comprising contacting a functional compound of the hBNPI protein with said substance, monitoring binding activity by physically detectable means, and identifying those substances which effect a chosen response. Preferably, the physically detectable means is competition with labeled inorganic phosphate or binding of ligand in an oocyte transient expression system

The instant invention provides such a screening system useful for discovering agents which compete with inorganic phosphate for binding to the hBNPI protein, said screening system comprising the steps of:

- a) isolating a human hBNPI protein;
- b) exposing said human hBNPI protein to a potential inhibitor or surrogate of the P_i/hBNPI protein complex;
- 25 c) introducing P_i;
 - d) removing non-specifically bound molecules; and
 - e) quantifying the concentration of bound potential inhibitor and/or P_i.
- 30 This allows one to rapidly screen for inhibitors or surrogates of the formation of the $P_i/hBNPI$ protein complex. Utilization of the screening system described above provides a sensitive and rapid means to determine compounds which

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interfere with the formation of the P_i/hBNPI protein complex. This screening system may also be adapted to automated procedures such as a PANDEX® (Baxter-Dade Diagnostics) system allowing for efficient high-volume screening of potential therapeutic agents.

In such a screening protocol a hBNPI protein is prepared as elsewhere described herein, preferably using recombinant DNA technology. A sample of a test compound is then introduced to the reaction vessel containing the hBNPI protein followed by the addition of P_i . In the alternative the P_i may be added simultaneously with the test compound. Unbound molecules are washed free and the eluent inspected for the presence of P_i or the test compound.

For example, in a preferred method of the 15 invention, radioactively labeled Pi may be used. The eluent is then scored for the radioactivity. The absence or diminution of the chemical label or radioactivity indicates the formation of the P_i/hBNPI protein complex. indicates that the test compound has not effectively competed 20 with P_i in the formation of the P_i/hBNPI protein complex. presence of the chemical label or radioactivity indicates that the test compound has competed with P_{i} in the formation of the P_i/hBNPI protein complex. Similarly, a radioactively or chemically labeled test compound may be used in which case the same steps as outlined above would be used except that 25 the interpretation of results would be the converse of using radioactively labelled Pi.

As would be understood by the skilled artisan these assays may also be performed such that the practitioner measures the radioactivity remaining with the protein, not in the eluent. A preferred such assay employs radiolabeled P_i. After the competition reaction has been performed the reaction mixture is then passed through a filter, the filter retaining the receptor and whatever is complexed with the receptor. The radioactivity on each filter is then measured in a scintillation counter. In such an assay higher amounts

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of radiolabel present indicate lower affinity for the receptor by the test compound.

The hBNPI protein may be free in solution or bound to a solid support. Whether the hBNPI protein is bound to a support or is free in solution, it is generally important that the conformation of the protein be conserved. In a preferred practice of the invention, therefore, the hBNPI protein is suspended in a hydrophobic environment employing natural or synthetic detergents, membrane suspensions, and the like. Preferred detergent complexes include the zwitterionic detergent 3-[(3-cholamidopropyl)-dimethylammonio]-1-propane sulfonate ("CHAPS") as well as sodium deoxycholate.

Skilled artisans will recognize that desirable dissociation constant (K_i) values are dependent on the selectivity of the compound tested. For example, a compound with a K_i which is less than 10 nM is generally considered an excellent candidate for drug therapy. However, a compound which has a lower affinity, but is selective for the particular receptor, may be an even better candidate. The present invention, however, provides radiolabeled competition assays, whether results therefrom indicate high affinity or low affinity to hBNPI protein, because skilled artisans will recognize that any information regarding binding or selectivity of a particular compound is beneficial in the pharmaceutical development of drugs.

Assays useful for evaluating ion channel cotransporters are well known in the art. <u>See. e.g.</u>, B. Ni, <u>et al.</u>, <u>supra</u>. One such assay is described below.

Functional analysis of hBNPI in transfected COS-1 cells

To confirm the functional properties of the hBNPI protein, we constructed the hBNPI cDNA into a mammalian expression vector (pcDNA3) and transfected the pcDNA3-hBNPI constructs into the COS-1 cells. Sodium-dependent ³²Pi uptake

in the cells transfected with hBNPI was stimulated 2-3 fold above that of those transfected with vectors alone or of nontransfected cells. Replacement of sodium chloride with choline chloride reduced ³²Pi uptake to background levels. Northern blot analysis was employed to examine the expression of hBNPI gene in transfected COS-1 cell lines. Labeled hBNPI cDNA detected strong expression of hBNPI transcripts in the COS-1 cells transfected with hBNPI but not in those cells transfected with the vector alone.

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Expression of hBNPI mRNA in human brain

We examined hBNPI expression in multiple human tissues by probing polyadenylated RNA from heart, brain, placenta, lung, liver, skeletal muscle, kidney, pancreas, spleen, thymus, prostate, testis, ovary, small intestine, colon and peripheral blood leukocytes. The Northern blot analysis demonstrated that hBNPI probe detected a single mRNA species of 2.8 kb and strong expression of hBNPI transcript 20 in the brain tissue. Trace levels of the hBNPI could be detected in RNA fractions from the small intestine, colon and testis if the blot was overexposured for a longer period of time (five days versus the usual one day exposure). signal could be detected in the other tissues. The level of 25 hBNPI in the brain fraction is at least 100 times higher than that in the intestine or colon. Northern blot analysis with multiple human brain regions shows that hBNPI mRNA is expressed in specific brain regions: most abundantly in neuron-enriched areas such as the amygdala and hippocampus; 30 at moderate levels in glia-enriched areas such as the corpus callosum; and at low levels in the substantia niga, subthalamic nuclei and thalamus. No hBNPI transcript was detected in RNAs isolated from the caudate nucleus and hypothalamus.

A Northern blot of human brain mRNA isolated from fetal and adult (37 yr-old) brain was prepared for the characterization of expression of the hBNPI during brain

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development. The blot was hybridized with $^{32}\text{P-labeled}$ hBNPI cDNA and human β -actin cDNA. The relative abundance of hBNPI mRNA shows a dramatic increase during postnatal development.

In situ hybridization histochemistry was employed to examine cells which express hBNPI transcripts in the human brain. hBNPI mRNA is highly expressed in the hippocampus formation and cerebral cortex. While the hybridization signal is present in various layers of the cerebral cortex, it appears to be more abundant in the neuronal layer v-vi where a distinct labeling is observed of pyramidal and nonpyramidal neurons. On closer inspection, it is apparent that hBNPI transcripts are concentrated in the pyramidal neurons of hippocampus and granule neurons of dentate gyrus. The hybridization signal was also detected in glia-enriched areas such as the corpus callosum, a finding which is consistent with data observed in Northern blot analysis of hBNPI mRNA in the human brain, and which suggests that, unlike its rat counterpart rBNPI, the hBNPI mRNA is expressed not only in neurons but also in glia as well. Cf., Ni, et al., supra.

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Genomic analysis of the hBNPI gene

Genomic Southern blotting is a valuable tool for identifying homologous genes in various species. We used hBNPI cDNA to detect hBNPI genes in a variety of vertebrate species under stringent hybridization condition. The species tested included human, monkey, rat, mouse, dog, cow, rabbit, chicken and yeast. One major fragment which appears to harbor hBNPI gene was detected in the human, monkey, dog, cow and rabbit. Two fragments generated by internal EcoRI sites were detected in the rat and mouse. No signal was detected in yeast DNA. The results suggest that hBNPI sequence is well conserved among vertebrate species.

Genomic DNAs derived from four human individuals

were digested with restriction endonuclases and used to
determine the hBNPI gene structure and possible polymorphisms
by Southern blot techniqus utilizing the full length hBNPI

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cDNA as a probe. The restriction patterns derived from 9 restrictions endonuclases are rather simple, and are similar between the four individuals. One major hybridizing fragment is generated by internal EcoRI, BalII, HindIII, PstI, PvuII, respectively. One major fragment with multiple weak hybridizing bands was generated by internal digestion with TaqI, MspI and BamHI. The results suggest that hBNPI gene structure is compact, that it is most likely present as a single copy, and that no polymorphisms of hBNPI gene exist.

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Chromosome localization

Using hBNPI cDNA we screened a library constructed with human leukocyte DNA to isolate the hBNPI gene. After 15 several rounds of screening, a 23 kb DNA fragment was isolated and identified as hBNPI gene. The hBNPI gene was labeled with digoxigenin dUTP by nick translation and hybridized to normal metaphase chromosomes derived from PHAstimulated peripheral blood lymphocytes using a fluorescent 20 in situ hybridization (FISH) technique. A specific hybridization signal was detected in the long arm of chromosome 19. Assignment of the hBNPI gene to the region of 19 was further confirmed by colocalization of a chromosome 19 specific probe, E2A, with the hBNPI gene. Measurements of ten specifically hybridized chromosomes 19 demonstrated that 25 hBNPI gene is located 66% of the distance from the centromere to the telomere of chromosome arm 19q, an area that corresponds to band 19q13.3. No positive signals were observed in any other chromosomes. Analysis of interphase 30 cells show only one copy of the probe present in the human genome, a finding which is consistent with the results of the genomic Southern blot.

The previously described screening systems identify compounds which competitively bind to the hBNPI protein.

Determination of the ability of such compounds to stimulate or inhibit the action of the hBNPI protein is essential to

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further development of such compounds for therapeutic applications. The need for a bioactivity assay system which determines the response of the hBNPI protein to a compound is clear. The instant invention provides such a bioactivity assay, said assay comprising the steps of:

- a) transfecting a mammalian host cell with an expression vector comprising DNA encoding a hBNPI protein;
- b) culturing said host cell under conditions such that the DNA encoding the hBNPI protein is expressed,
- c) exposing said host cell so transfected to a test compound, and
 - d) measuring the change in a physiological condition known to be influenced by the binding of a cation to the hBNPI protein relative to a control in which the transfected host cell is not exposed to the test compound.

An oocyte transient expression system can be constructed according to the procedure described in S. Lübbert, et al., Proceedings of the National Academy of Sciences (USA), 84:4332 (1987).

In an especially preferred embodiment of this invention an assay measuring the inhibition of radiolabeled phosphate uptake was performed. The inhibition of phosphate uptake is a relatively simple assay used to determine these agents which negatively affect the proteins of the present invention.

In another embodiment this invention provides a

35 method for identifying, in a test sample, DNA homologous to a
probe of the present invention, wherein the test nucleic acid
is contacted with the probe under hybridizing conditions and

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identified as being homologous to the probe. Hybridization techniques are well known in the art. See. e.g., J. Sambrook, et al., supra, at Chapter 11.

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The nucleic acid compounds of the present invention may also be used to hybridize to genomic DNA which has been digested with one or more restriction enzymes and run on an electrophoretic gel. The hybridization of radiolabeled probes onto such restricted DNA, usually fixed to a membrane after electrophoresis, is well known in the art. See, e.g., J. Sambrook, supra. Such procedures may be employed in searching for persons with mutations in these receptors by the well-known techniques of restriction fragment length polymorphisms (RFLP), the procedures of which are described in U.S. Patent 4,666,828, issued May 19, 1987, the entire contents of which is herein incorporated by reference.

The proteins of this invention as well as fragments of these proteins may be used as antigens for the synthesis of antibodies. The term "antibody" as used herein describes antibodies, fragments of antibodies (such as, but not limited, to Fab, Fab', Fab2', and Fv fragments), and chimeric, humanized, veneered, resurfaced, or CDR-grafted antibodies capable of binding antigens of a similar nature as the parent antibody molecule from which they are derived. The instant invention also encompasses single chain polypeptide binding molecules.

The term "antibody" as used herein is not limited by the manner in which the antibodies are produced, whether such production is in situ or not. The term "antibody" as used in this specification encompasses those antibodies produced by recombinant DNA technology means including, but not limited, to expression in bacteria, yeast, insect cell lines, or mammalian cell lines.

The production of antibodies, both monoclonal and polyclonal, in animals, especially mice, is well known in the art. See. e.g., C. Milstein, HANDBOOK OF EXPERIMENTAL IMMUNOLOGY, (Blackwell Scientific Pub., 1986); J. Goding, Monoclonal ANTIBODIES: PRINCIPLES AND PRACTICE, (Academic Press, 1983). For

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the production of monoclonal antibodies the basic process begins with injecting a mouse, or other suitable animal, with an immunogen. The mouse is subsequently sacrificed and cells taken from its spleen are fused with myeloma cells, resulting in a hybridoma that reproduces in vitro. The population of hybridomas is screened to isolate individual clones, each of which secretes a single antibody species, specific for the immunogen. The individual antibody species obtained in this way is each the product of a single B cell from the immune animal generated in response to a specific antigenic site, or epitope, recognized on the immunogenic substance.

Chimeric antibodies are described in U.S. Patent No. 4,816,567, which issued March 28, 1989 to S. Cabilly, et This reference discloses methods and vectors for the preparation of chimeric antibodies. The entire contents of U.S. Patent No. 4,816,567 are herein incorporated by reference. An alternative approach to production of genetically engineered antibodies is provided in U.S. Patent No. 4,816,397, which also issued March 28, 1989 to M. Boss, 20 et al., the entire contents of which are herein incorporated by reference. The Boss patent teaches the simultaneous coexpression of the heavy and light chains of the antibody in the same host cell.

The approach of U.S. Patent 4,816,397 has been 25 further refined as taught in European Patent Publication No. 0 239 400, which published September 30, 1987. The teachings of this European patent publication (Winter) are a preferred format for the genetic engineering of the reactive monoclonal antibodies of this invention. The Winter technology involves 30 the replacement of complementarity determining regions (CDRs) of a human antibody with the CDRs of a murine monoclonal antibody thereby converting the specificity of the human antibody to the specificity of the murine antibody which was the source of the CDR regions. This "CDR grafting" 35 technology affords a molecule containing minimal murine sequence and thus is less immunogenic.

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Single chain antibody technology is yet another variety of genetically engineered antibody which is now well known in the art. See, e.g. R.E. Bird, et al., Science 242:423-426 (1988); Patent Cooperation Treaty Publication No. WO 88/01649, which was published 10 March 1988. The single chain antibody technology involves joining the binding regions of heavy and light chains with a polypeptide sequence to generate a single polypeptide having the binding specificity of the antibody from which it was derived.

The aforementioned genetic engineering approaches provide the skilled artisan with numerous means to generate molecules which retain the binding characteristics of the parental antibody while affording a less immunogenic format.

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These antibodies are used in diagnostics, 15 therapeutics or in diagnostic/therapeutic combinations. "diagnostics" as used herein is meant testing that is related to either the in vitro or in vivo diagnosis of disease states or biological status in mammals, preferably in humans. By "therapeutics" and "therapeutic/diagnostic combinations" as 20 used herein is respectively meant the treatment or the diagnosis and treatment of disease states or biological status by the in vivo administration to mammals, preferably humans, of the antibodies of the present invention. antibodies of the present invention are especially preferred 25 in the diagnosis and/or treatment of conditions associated with an excess or deficiency of hBNPI proteins.

In addition to being functional as direct therapeutic and diagnostic aids, the availability of a family of antibodies which are specific for the hBNPI protein enables the development of numerous assay systems for detecting agents which bind to this protein. One such assay system comprises radiolabeling hBNPI protein-specific antibodies with a radionuclide such as ¹²⁵I and measuring displacement of the radiolabeled hBNPI protein-specific antibody from solid phase hBNPI protein in the presence of a potential antagonist or inhibitor.

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Numerous other assay systems are also readily adaptable to detect agents which bind hBNPI protein. Examples of these aforementioned assay systems are discussed in Methods in Enzymology, (J. Langone. and H. Vunakis, eds. 1981), Vol. 73, Part B, the contents of which are herein incorporated by reference. Skilled artisans are directed to Section II of Methods in Enzymology, Vol. 73, Part B, supra, which discusses labeling of antibodies and antigens, and

In addition to the aforementioned antibodies specific for the hBNPI protein, this invention also provides antibodies which are specific for the hypervariable regions of the anti-hBNPI protein antibodies. Some such anti-idiotypic antibodies would resemble the original epitope, the hBNPI protein, and, therefore, would be useful in evaluating the effectiveness of compounds which are potential antagonists, agonists, or partial agonists of the hBNPI protein. See, e.g., Cleveland, et al., Nature (London), 305:56 (1983); Wasserman, et al., Proceedings of the National Academy of Sciences (USA), 79:4810 (1982).

Section IV, which discusses immunoassay methods.

In another embodiment, this invention encompasses pharmaceutical formulations for parenteral administration which contain, as the active ingredient, the anti-hBNPI protein antibodies described, <u>supra</u>. Such formulations are prepared by methods commonly used in pharmaceutical chemistry.

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Products for parenteral administration are often formulated and distributed in solid, preferably freeze-dried form, for reconstitution immediately before use. Such formulations are useful compositions of the present invention. Their preparation is well understood by pharmaceutical chemists.

In general, these formulations comprise the active ingredient in combination with a mixture of inorganic salts, to confer isotonicity, as well as dispersing agents such as lactose, to allow the dried preparation to dissolve quickly

upon reconstitution. Such formulations are reconstituted for use with highly purified water to a known concentration.

Alternatively, a water soluble form of the antibody can be dissolved in one of the commonly used intravenous fluids and administered by infusion. Such fluids include physiological saline, Ringer's solution or a 5% dextrose solution.

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Ni, Binhui
 Paul, Steven M.
 - (ii) TITLE OF INVENTION: HUMAN BRAIN SODIUM DEPENDENT INORGANIC PHOSPATE COTRANSPROTER AND RELATED NUCLEIC ACID COMPOUNDS
 - (iii) NUMBER OF SEQUENCES: 3
 - (iv) CORRESPONDENCE ADDRESS:
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 - (B) STREET: Lilly Corporate Center
 - (C) CITY: Indianapolis
 - (D) STATE: Indiana
 - (E) COUNTRY: United States of America
 - (F) ZIP: 46285
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
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- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2716 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 461..2143

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CGATAAGCTT GATATCGAAT TCCGGACTCT TGCTCGGGCG CCTTAACCCG GCC	STTCGGTT 60
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CCGGCTCCAC GCCAGCGAGC CGGGCTTCTT ACCCATTTAA AGTTTGAGAA TAC	GCTTGAGA 180
TCGTTTCGGC CCCAAGACCT CTAATCATTC GCTTTACCGG ATAAAACTGC GTC	GCGGGGG 240
TGCGTCGGGT CTGCGAGAGC GCCAGCTATC CTGAGGGAAA CTTCGGAGGG AAC	CAGCTAC 300
TAGATGGTTC GATTAGTCTT TCGCCCCTAT ACCCAGGTCG GACGACCGAT TTC	GCACGTCA 360
GGACCGCTAC GGACCTCCAC CAGAGTTTCC TCTGGCTTCG CCCTGCCCAG GCG	SATCGGCG 420
GGGGGGACCC GCGGGGTGAC CGGCGGCAGG AGCCGCCACC ATG GAG TTC CG	
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Glu Glu Phe Arg Lys Leu Ala Gly Arg Ala Leu Gly Lys Leu Hi 10 15 2	s Arg 10
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GTG GTG GTG CAG AAA GCC CAG TTC AGC TGG GAT CCA GAG ACT GT	C GGC 811
Val Val Val Gln Lys Ala Gln Phe Ser Trp Asp Pro Glu Thr Va	
105 110 115	
CTC ATA CAC GGC TCC TTT TTC TGG GGC TAC ATT GTC ACT CAG AT	T CCA 859
Leu Ile His Gly Ser Phe Phe Trp Gly Tyr Ile Val Thr Gln Il	e Pro
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					TAC Tyr											1051
					CGG Arg											1099
					GTG Val											1147
					AGC Ser 235											1195
					TTC Phe											1243
					TCG Ser											1291
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					TTC Phe											1387
					AGC Ser 315											1435
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					CCC Pro											1531

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ACC	AAC		CGC	AAG	TTG	ATG	365 AAC	TGC	CCA	GGC	TTC	370 GGC	ΔΤΩ	GAA	GCC	1	627
											Phe 385					•	027
											AAG Lys					1	675
											TTC Phe					1	723
											TAC Tyr					1'	771
											GGC Gly					:	1819
											CGG Arg 465					:	1867
											GGA Gly					:	1915
											TGG Trp					1	1963
											CAT His					2	2011
											GAG Glu					2	2059
											CAC His 545					2	2107
	CCC Pro										TGA *	CCAT	GTGC	CT		2	153
CCC	CTGA	LAT G	GCAG	TTTC	C AG	GACC	TCCA	TTC	CACI	CAT	СТСТ	GGCC	TG A	GTGA	CAGTG	2	213
TCAZ	GGAZ	cc c	TGCI	CCTC	T CI	GTCC	TGCC	TCA	GCC	TAA	GAAG	CACI	CT C	CCTI	GTTCC	2	273

CAGTY	GCTG	TC A	YAAA	CCTC	T T	CCTT	CCCA	A TT	GCCT	CTCA	GGG	GTAG	TGA	AGCT	GCAGAC
TGAC	AGTT	TC I	AAGG/	ATAC	CC A)TTAA	cccc	r aa	AGGT	TCCC	TCT	CCAC	CCG	TTCT	GCCTCA
GTGG:	TTTC	AA A	ATCT	CTÇC	rt t	CAGG	GCTT!	T AT	TTGA	atgg	ACA	GTTC	GAC	CTCT	ТАСТСТ
CTCT	TGTG	GT 1	MTG	AGGC	AC C	CACA	cccc	CG	CTTT	CCTT	TAT	CTCC	AGG	GACT	CTCAGG
CTAA	CCTT	TG A	AGAT	CACTO	CA G	CTCC	CATC	r cc	TTTC	AGAA	AAA	TTCA	AGG	TCCT	ССТСТА
GAAG?	TTTC.	AA 1	itct (CTCC	CA A	CTCT	STTC	r gc	ATCT	TCCA	GAT	TGGT	TTA .	ACCA	ATTACT
CGTC	CCCG	CC A	ATTCO	CAGG	GA T	IGAT.	rctc <i>i</i>	A CC	AGCG'	TTTC	TGA	TGGA	AAA '	TGGC	GGGAAT
TCCTC	GCAG	cc c	CGGG	GAT	CC A	CT									
(2)	INFO	RMAT	rion	FOR	SEQ	ID I	NO:2:	:							
	(i) S	EOUI	ENCE	CHA	RACTI	ERIST	rics	:	٠					
	•		(A)) LEI	NGTH	: 56	l ami	ino a		s					
				TYI											
	(i	i) N	OLE	CULE	TYP	E: p	rotei	in							
	(x	i) S	EQUI	ENCE	DES	CRIP	rion:	: SE	Q ID	NO:2	2:				
Met (Glu	Phe	Arg	Gln	Glu	Glu	Phe	Arg	Lys	Leu	Ala	Gly	Arg	Ala	Leu
1				5					10			_		15	
Gly I	Lys :	Leu	His 20	Arg	Leu	Leu	Glu	Lys 25	Arg	Gln	Glu	Gly	Ala 30	Glu	Thr
Leu C	Glu :	Leu 35	Ser	Ala	Asp	Gly	Arg 40	Pro	Val	Thr	Thr	Gln 45	Thr	Arg	Asp
Pro I	Pro '	Val	Val	Asp	_	Thr 55	_	Phe	Gly	Leu	Pro 60	_	Arg	Tyr	Ile
Ile A	Ala	Ile	Met	Ser	Gly 70	Leu	Gly	Phe	Суз	Ile 75	Ser	Phe	Gly	Ile	Arg 80
Cys /	Asn :	Leu	Gly	Val 85	Ala	Ile	Val	Ser	Met 90	Val	Asn	Asn	Ser	Thr 95	Thr
His A	Arg (Gly	Gly 100	His	Val	Val	Val	Gln 105	Lys	Ala	Gln	Phe	Ser 110	Trp	Asp
Pro C		Thr 115	Val	Gly	Leu	Ile	His 120	Gly	Ser	Phe	Phe	Trp 125	Gly	Tyr	Ile
Val 7	Thr (Gln	Ile	Pro	Gly	Gly	Phe	Ile	Cys	Gln	Lys	Phe	Ala	Ala	Asn

	Arg 145	Val	Phe	Gly	Phe	Ala 150	Ile	Val	Ala	Thr	Ser 155	Thr	Leu	Asn	Met	Leu 160
	Ile	Pro	Ser	Ala	Ala 165	Arg	Val	His	Tyr	Gly 170	Сув	Val	Ile	Phe	Val 175	Arg
	Ile	Leu	Gln	Gly 180	Leu	Val	Glu	Gly	Val 185	Thr	Tyr	Pro	Ala	Cys 190	His	Gly
	Ile	Trp	Ser 195	Lys	Trp	Ala	Pro	Pro 200	Leu	Glu	Arg	Ser	Arg 205	Leu	Ala	Thr
1	Thr	Ala 210	Phe	Cys	Gly	Ser	Tyr 215	Ala	Gly	Ala	Val	Val 220	Ala	Met	Pro	Leu
	Ala 225	Gly	Val	Leu	Val	Gln 230	Tyr	Ser	Gly	Trp	Ser 235	Ser	Val	Phe	Tyr	Val 240
ı	Tyr	Gly	Ser	Phe	Gly 245	Ile	Phe	Trp	Tyr	Leu 250	Phe	Trp	Leu	Leu	Val 255	Ser
:	Tyr	Glu	Ser	Pro 260	Ala	Leu	His	Pro	Ser 265	Ile	Ser	Glu	Glu	Glu 270	Arg	Lys
1	Tyr	Ile	Glu 275	Asp	Ala	Ile	Gly	Glu 280	Ser	Ala	Lys	Leu	Met 285	Asn	Pro	Leu
1	Thr	Lys 290	Phe	Ser	Thr	Pro	Trp 295	Arg	Arg	Phe	Phe	Thr 300	Ser	Met	Pro	Val
	Tyr 305	Ala	Iļe	Ile	Val	Ala 310	Asn	Phe	Сув	Arg	Ser 315	Trp	Thr	Phe	Tyr	Leu 320
,	Leu	Leu	Ile	Ser	Gln 325	Pro	Asp	Tyr	Phe	Glu 330	Glu	Val	Phe	Gly	Phe 335	Glu
	Iļe	Ser	Lys	Val 340	Gly	Leu	Val	Ser	Ala 345	Leu	Pro	His	Leu	Val 350	Met	Thr
	Ile	Ile	Val 355	Pro	Ile	Gly	Gly	Gln 360	Ile	Ala	Asp	Phe	Leu 365	Arg	Ser	Arg
	Arg	Ile 370	Met	Ser	Thr	Thr	Asn 375	Val	Arg	Lys	Leu	Met 380	Asn	Cys	Gly	Gly
	Phe 385	Gly	Met	Glu	Ala	Thr 390	Leu	Leu	Leu	Val	Val 395	Gly	Tyr	Ser	His	Ser 400
,	Lys	Gly	Val	Ala	Ile 405	Ser	Phe	Leu	Val	Leu 410	Ala	Val	Gly	Phe	Ser 415	Gly
	Phe	Ala	Ile	Ser 420	Gly	Phe	Asn	Val	Asn 425	His	Leu	yab	Ile	Ala 430	Pro	Arg

lyr	MIG	435	116	Leu	met	GIŞ	440	ser	ASI	GIY	vai	445	Inr	Dea	Ser
Gly	Met 4 50	Val	Cys	Pro	Ile	11e 455	Val	Gly	Ala	Met	Thr 460	Lys	His	Lys	Thr
Arg 465	Glu	Glu	Trp	Gln	Tyr 470	Val	Phe	Leu	Ile	Ala 475	Ser	Leu	Val	His	Tyr 480
Gly	Gly	Val	Ile	Phe 485	Tyr	Gly	Val	Phe	Ala 490	Ser	Gly	Glu	Lys	Gln 495	Pro
Trp	Ala	Glu	Pro 500	Glu	Glu	Met	Ser	Glu 505	Glu	Lys	Cys	Gly	Phe 510	Val	Gly
His	Asp	Gln 515	Leu	Ala	Gly	Ser	Asp 520	Asp	Ser	Glu	Met	Glu 525	Asp	Glu	Ala
Glu	Pro 530		Gly	Ala	Pro	Pro 535	Ala	Pro	Pro	Pro	Ser 540	Tyr	Gly	Ala	Thr
His 545			Phe	Gln			Arg			Pro		Val	_	_	Tyr 560

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2716 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

60	GCGUUCGGUU	CCUUAACCCG	UGCUCGGGCG	UCCGGACUCU	GAUAUCGAAU	CGAUAAGCUU
120	CAUUCCACGC	UAGGCACUCG	AGUGGCCCAC	GCUUACCAAA	CGCCAGUUCU	CAUCCCGCAG
180	UAGGUUGAGA	AGUUUGAGAA	ACCCAUUUAA	CGGGCUUCUU	GCCAGCGAGC	CCGGCUCCAC
240	GUGGCGGGG	AUAAAACUGC	GCUUUACCGG	CUAAUCAUUC	CCCAAGACCU	UCGUUUCGGC
300	AACCAGCUAC	CUUCGGAGGG	CUGAGGGAAA	GCCAGCUAUC	CUGCGAGAGC	UGCGUCGGGU
360	UUGCACGUCA	GACGACCGAU	ACCCAGGUCG	UCGCCCCUAU	GAUUAGUCUU	UAGAUGGUUC
420	GCGAUCGGCG	CCCUGCCCAG	UCUGGCUUCG	CAGAGUUUCC	GGACCUCCAC	GGACCGCUAC

GGGGGACCC	GCGGGGUGAC	CGGCGGCAGG	AGCCGCCACC	AUGGAGUUCC	GCCAGGAGGA	480
GUUUCGGAAG	CUAGCGGGUC	GUGCUCUCGG	GAAGCUGCAC	CGCCUUCUGG	AGAAGCGGCA	540
GGAAGGCGCG	GAGACGCUGG	AGCUGAGUGC	GGAUGGGCGC	CCGGUGACCA	CGCAGACCCG	600
GGACCCGCCG	GUGGUGGACU	GCACCUGCUU	cgccucccu	CGCCGCUACA	UUAUCGCCAU	660
CAUGAGUGGU	CUGGGCUUCU	GCAUCAGCUU	UGGCAUCCGC	UGCAACCUGG	GCGUGGCCAU	720
CGUCUCCAUG	GUCAAUAACA	GCACGACCCA	CCGCGGGGGC	CACGUGGUGG	UGCAGAAAGC	780
CCAGUUCAGC	UGGGAUCCAG	AGACUGUCGG	CCUCAUACAC	GGCUCCUUUU	UCUGGGGCUA	840
CAUUGUCACU	CAGAUUCCAG	GAGGAUUUAU	CUGUCAAAAA	UUUGCAGCCA	ACAGAGUUUU	900
CGGCUUUGCU	AUUGUGGCAA	CAUCCACUCU	AAACAUGCUG	AUCCCCUCAG	cugcccgcgu	960
CCACUAUGGC	UGUGUCAUCU	UCGUGAGGAU	CCUGCAGGGG	UUGGUAGAGG	GGGUCACAUA	1020
cccccccucc	CAUGGGAUCU	GGAGCAAAUG	GGCCCCACCC	UUAGAACGGA	GUCGCCUGGC	1080
GACGACAGCC	UUUUGUGGUU	CCUAUGCUGG	GGCGGUGGUC	GCGAUGCCCC	UCGCCGGGGU	1140
CCUUGUGCAG	UACUCAGGAU	GGAGCUCUGU	UUUCUACGUC	UACGGCAGCU	UCGGGAUCUU	1200
CUGGUACCUG	UUCUGGCUGC	UCGUCUCCUA	CGAGUCCCCC	GCGCUGCACC	CCAGCAUCUC	1260
GGAGGAGGAG	CGCAAGUACA	UCGAGGACGC	CAUCGGAGAG	AGCGCGAAAC	UCAUGAACCC	1320
CCUCACGAAG	UUUAGCACUC	CCUGGCGGCG	CUUCUUCACG	UCUAUGCCAG	UCUAUGCCAU	1380
CAUCGUGGCC	AACUUCUGCC	GCAGCUGGAC	GUUCUACCUG	CUGCUCAUCU	CCCAGCCCGA	1440
CUACUUCGAA	GAAGUGUUCG	GCUUCGAGAU	CAGCAAGGUA	GGCCUGGUGU	CCGCGCUGCC	1500
CCACCUGGUC	AUGACCAUCA	UCGUGCCCAU	CGGCGGCCAG	AUCGCGGACU	UCCUGCGGAG	1560
CCGCCGCAUC	AUGUCCACCA	CCAACGUGCG	CAAGUUGAUG	AACUGCGGAG	GCUUCGGCAU	1620
GGAAGCCACG	CUGCUGUUGG	UGGUCGGCUA	CUCGCACUCC	AAGGGCGUGG	CCAUCUCCUU	1680
CCUGGUCCUA	GCCGUGGGCU	UCAGCGGCUU	CGCCAUCUCU	GGGUUCAACG	UGAACCACCU	1740
GGACAUAGCC	CCGCGCUACG	CCAGCAUCCU	CAUGGGCAUC	UCCAACGGCG	UGGGCACACU	1800
GUCGGGCAUG	GUGUGCCCCA	UCAUCGUGGG	GGCCAUGACU	AAGCACAAGA	CUCGGGAGGA	1860
GUGGCAGUAC	GUGUUCCUAA	uugccucccu	GGUGCACUAU	GGAGGUGUCA	UCUUCUACGG	1920
GGUCUUUGCU	UCUGGAGAGA	AGCAGCCGUG	GGCAGAGCCU	GAGGAGAUGA	GCGAGGAGAA	1980
GUGUGGCUUC	GUUGGCCAUG	ACCAGCUGGC	UGGCAGUGAC	GACAGCGAAA	UGGAGGAUGA	2040

GGCUGAGCCC	CCGGGGGCAC	CCCCUGCACC	CCCGCCCUCC	UAUGGGCCA	CACACAGCAC	2100
AUUUCAGCCC	CCCAGGCCCC	CACCCCCUGU	CCGGGACUAC	UGACCAUGUG	CCUCCCACUG	2160
AAUGGCAGUU	UCCAGGACCU	CCAUUCCACU	CAUCUCUGGC	CUGAGUGACA	GUGUCAAGGA	2220
ACCCUGCUCC	ucucuguccu	GCCUCAGGCC	UAAGAAGCAC	ucucccuugu	UCCCAGUGCU	2280
GUCAAAUCCU	cuuuccuucc	CAAUUGCCUC	UCAGGGGUAG	UGAAGCUGCA	GACUGACAGU	2340
UUCAAGGAUA	CCCAAAUUCC	CCUAAAGGUU	CCCUCUCCAC	ccguucugcc	UCAGUGGUUU	2400
CAAAUCUCUC	CUUUCAGGGC	UUUAUUUGAA	UGGACAGUUC	GACCUCUUAC	ucucucuugu	2460
GGUUUUGAGG	CACCCACACC	ccccccuuuc	CUUUAUCUCC	AGGGACUCUC	AGGCUAACCU	2520
UUGAGAUCAC	UCAGCUCCCA	UCUCCUUUCA	GAAAAAUUCA	AGGUCCUCCU	CUAGAAGUUU	2580
CAAAUCUCUC	CCAACUCUGU	UCUGCAUCUU	CCAGAUUGGU	UUAACCAAUU	ACUCGUCCCC	2640
GCCAUUCCAG	GGAUUGAUUC	UCACCAGCGU	UUCUGAUGGA	AAAUGGCGGG	AAUUCCUGCA	2700
GCCCGGGGA	UCCACU					2716

We Claim:

 An isolated amino acid compound functional as a human brain Na⁺-dependent inorganic phosphate cotransporter
 which comprises the amino acid sequence

	Met 1		Phe	Arg	Gln 5	Glu	Glu	Phe	Arg	Lys 10		Ala	Gly	Arg	Ala 15	
10	Gly	Lys	Leu	His 20	Arg	Leu	Leu	Glu	Lys 25		Gln	Glu	Gly	Ala 30		Thr
15	Leu	Glu	Leu 35	Ser	Ala	Asp	Gly	Arg 40	Pro	Val	Thr	Thr	Gln 45	Thr	Arg	Asp
	Pro	Pro 50	Val	Val	Asp	Cys	Thr 55	Cys	Phe	Gly	Leu	Pro 60		Arg	Tyr	Ilė
20	Ile 65	Ala	Ile	Met	Ser	Gly 70	Leu	Gly	Phe	Суз	Ile 75	Ser	Phe	Gly	Ile	Arg 80
	Cys	Asn	Leu	Gly	Val 85	Ala	Ile	Val	Ser	Met 90	Val	Asn	Asn	Ser	Thr 95	Thr
25	His	Arg	Gly	Gly 100	His	Val	Val	Val	Gln 105	Lys	Ala	Gln	Phe	Ser 110	Trp	Asp
30	Pro	Glu	Thr 115	Val	Gly	Leu	Ile	His 120	Gly	Ser	Phe	Phe	Trp 125	Gly	Tyr	Ile
	Val	Thr 130	Gln	Ile	Pro	Gly	Gly 135	Phe	Ile	Сув	Gln	Lys 140	Phe	Ala	Ala	Asn
35	Arg 145	Val	Phe	Gly	Phe	Ala 150	Ile	Val	Ala	Thr	Ser 155	Thr	Leu	Asn	Met	Leu 160
	Ile	Pro	Ser	Ala	Ala 165	Arg	Val	His	Tyr	Gly 170	Cys	Val	Ile	Phe	Val 175	Arg
40	Ile	Leu	Gln	Gly 180	Leu	Val	Glu	Gly	Val 185	Thr	Tyr	Pro	Ala	Суз 190	His	Gly
45	Ile	Trp	Ser 195	Lys	Trp	Ala	Pro	Pro 200	Leu	Glu	Arg	Ser	Arg 205	Leu	Ala	Thr
	Thr	Ala 210	Phe	Cys	Gly	Ser	Tyr 215	Ala	Gly	Ala	Val	Val 220	Ala	Met	Pro	Leu
50	Ala 225	Gly	Val	Leu		Gln 230	Tyr	Ser	Gly	Trp	Ser 235	Ser	Va1	Phe	Týr	Val 240

	•	Tyr	Gly	Ser	Phe	Gly 245	Ile	Phe	Trp	Tyr	Leu 250	Phe	Trp	Leu	Leu	Val 255	Ser
5	Ç	Tyr	Glu	Ser	Pro 260	Ala	Leu	His	Pro	Ser 265	Ile	Ser	Glu	Glu	Glu 270	Arg	Lys
	5	lyr	Ile	Glu 275	Asp	Ala	Ile	Gly	Glu 280	Ser	Ala	Lys	Leu	Met 285	Asn	Pro	Leu
10	5	Thr .	Lys 290	Phe	Ser	Thr	Pro	Trp 295	Arg	Arg	Phe	Phe	Thr 300	Ser	Met	Pro	Val
15		Tyr 305	Ala	Ile	Ile	Val	Ala 310	Asn	Phe	Cys	Arg	Ser 315	Trp	Thr	Phe	Tyr	Leu 320
	1	Leu	Leu	Ile	Ser	Gln 325	Pro	Asp	Tyr	Phe	Glu 330	Glu	Val	Phe	Gly	Phe 335	Glu
20	:	Ile	Ser	Lys	Val 340	Gly	Leu	Val	Ser	Ala 345	Leu	Pro	His	Leu	Val 350	Met	Thr
	•	Ile	Ile	Va1 355	Pro	Ile	Gly	Gly	Gln 360	Ile	Ala	Asp	Phe	Leu 365	Arg	Ser	Arg
25	2	Arg	Ile 370	Met	Ser	Thr	Thr	Asn 375	Val	Arg	Lys	Leu	Met 380	Asn	Cys	Gly	Gly
30		Phe 385	Gly	Met	Glu	Ala	Thr 390	Leu	Leu	Leu	Val	Va1 395	Gly	Tyr	Ser	His	Ser 400
	1	Lys	Gly	Val	Ala	Ile 405	Ser	Phe	Leu	Val	Leu 410	Ala	Val	Gly	Phe	Ser 415	Gly
35	1	Phe	Ala	Ile	Ser 420	Gly	Phe	Asn	Val	A sn 4 25	His	Leu	Asp	Ile	Ala 430	Pro	Arg
	1	Tyr	Ala	Ser 435	Ile	Leu	Met	Gly	Ile 440	Ser	Asn	Gly	Val	Gly 445	Thr	Leu	Ser
40	(Gly	Met 450	Val	Cys	Pro	Ile	11e 455	Val	Gly	Ala	Met	Thr 460	Lys	His	Lys	Thr
45		Arg 465	Glu	Glu	Trp	Gln	Tyr 470	Val	Phe	Leu	Ile	Ala 475	Ser	Leu	Val	His	Tyr 480
	(Gly	Gly	Val	Ile	Phe 485	Tyr	Gly	Val	Phe	Ala 490	Ser	Gly	Glu	Lys	Gln 495	Pro
50	5	Trp	Ala	Glu	Pro 500	Glu	Glu	Met	Ser	Glu 505	Glu	Lys	Cys	Gly	Phe 510	Val	Gly
	1	His	Asp	Gln 515	Leu	Ala	Gly	Ser	Asp 520	Asp	Ser	Glu	Met	Glu 525	Asp	Glu	Ala
55	C	Glu	Pro	Pro	Gly	Ala	Pro	Pro	Ala	Pro	Pro	Pro	Ser	Tyr	Gly	Ala	Thr

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57

530 535 540

His Ser Thr Phe Gln Pro Pro Arg Pro Pro Pro Pro Val Arg Asp Tyr 545 550 555 560

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which is SEQ ID NO:2, or a functional equivalent thereof, or a fragment of at least 6 continuous amino acids thereof.

- 2. A nucleic acid compound encoding an amino acid compound of Claim 1.
 - 3. A composition comprising an isolated nucleic acid compound containing a sequence encoding a human brain Na+-dependent inorganic phosphate cotransporter or fragment thereof as claimed in Claim 2, wherein said sequence encoding a human brain Na+-dependent inorganic phosphate cotransporter or fragment thereof is selected from the group consisting of:
- 20 (a) CGATAAGCTT GATATCGAAT TCCGGACTCT TGCTCGGGCG CCTTAACCCG GCGTTCGGTT CATCCCGCAG CGCCAGTTCT GCTTACCAAA AGTGGCCCAC TAGGCACTCG CATTCCACGC CCGGCTCCAC GCCAGCGAGC CGGGCTTCTT ACCCATTTAA AGTTTGAGAA TAGGTTGAGA 25 TCGTTTCGGC CCCAAGACCT CTAATCATTC GCTTTACCGG ATAAAACTGC GTGGCGGGGG TGCGTCGGGT CTGCGAGAGC GCCAGCTATC CTGAGGGAAA CTTCGGAGGG AACCAGCTAC 30 TAGATGGTTC GATTAGTCTT TCGCCCCTAT ACCCAGGTCG GACGACCGAT TTGCACGTCA GGACCGCTAC GGACCTCCAC CAGAGTTTCC TCTGGCTTCG CCCTGCCCAG GCGATCGGCG GGGGGGACCC GCGGGTGAC CGGCGGCAGG AGCCGCCACC ATGGAGTTCC GCCAGGAGGA 35 GTTTCGGAAG CTAGCGGGTC GTGCTCTCGG GAAGCTGCAC CGCCTTCTGG AGAAGCGGCA GGAAGGCGCG GAGACGCTGG AGCTGAGTGC GGATGGGCGC CCGGTGACCA CGCAGACCCG 40 GGACCCGCCG GTGGTGGACT GCACCTGCTT CGGCCTCCCT CGCCGCTACA TTATCGCCAT CATGAGTGGT CTGGGCTTCT GCATCAGCTT TGGCATCCGC TGCAACCTGG GCGTGGCCAT CGTCTCCATG GTCAATAACA GCACGACCCA CCGCGGGGGC CACGTGGTGG TGCAGAAAGC 45 CCAGTTCAGC TGGGATCCAG AGACTGTCGG CCTCATACAC GGCTCCTTTT TCTGGGGCTA CATTGTCACT CAGATTCCAG GAGGATTTAT CTGTCAAAAA TTTGCAGCCA ACAGAGTTTT

	CGGCTTTGCT	ATTGTGGCAA	CATCCACTCT	AAACATGCTG	ATCCCCTCAG	CTGCCCGCGT
5	CCACTATGGC	TGTGTCATCT	TCGTGAGGAT	CCTGCAGGGG	TTGGTAGAGG	GGGTCACATA
J	CCCCGCCTGC	CATGGGATCT	GGAGCAAATG	GGCCCCACCC	TTAGAACGGA	GTCGCCTGGC
	GACGACAGCC	TTTTGTGGTT	CCTATGCTGG	GGCGGTGGTC	GCGATGCCCC	TCGCCGGGGT
10	CCTTGTGCAG	TACTCAGGAT	GGAGCTCTGT	TTTCTACGTC	TACGGCAGCT	TCGGGATCTI
	CTGGTACCTG	TTCTGGCTGC	TCGTCTCCTA	CGAGTCCCCC	GCGCTGCACC	CCAGCATCTC
15	GGAGGAGGAG	CGCAAGTACA	TCGAGGACGC	CATCGGAGAG	AGCGCGAAAC	TCATGAACCC
	CCTCACGAAG	TTTAGCACTC	CCTGGCGGCG	CTTCTTCACG	TCTATGCCAG	TCTATGCCAT
	CATCGTGGCC	AACTTCTGCC	GCAGCTGGAC	GTTCTACCTG	CTGCTCATCT	CCCAGCCCGA
20	CTACTTCGAA	GAAGTGTTCG	GCTTCGAGAT	CAGCAAGGTA	GGCCTGGTGT	CCGCGCTGCC
	CCACCTGGTC	ATGACCATCA	TCGTGCCCAT	CGGCGGCCAG	ATCGCGGACT	TCCTGCGGAG
25	CCGCCGCATC	ATGTCCACCA	CCAACGTGCG	CAAGTTGATG	AACTGCGGAG	GCTTCGGCAT
	GGAAGCCACG	CTGCTGTTGG	TGGTCGGCTA	CTCGCACTCC	AAGGGCGTGG	CCATCTCCTT
	CCTGGTCCTA	GCCGTGGGCT	TCAGCGGCTT	CGCCATCTCT	GGGTTCAACG	TGAACCACCT
30	GGACATAGCC	CCGCGCTACG	CCAGCATCCT	CATGGGCATC	TCCAACGGCG	TGGGCACACT
	GTCGGGCATG	GTGTGCCCCA	TCATCGTGGG	GGCCATGACT	AAGCACAAGA	CTCGGGAGGA
35	GTGGCAGTAC	GTGTTCCTAA	TTGCCTCCCT	GGTGCACTAT	GGAGGTGTCA	TCTTCTACGG
	GGTCTTTGCT	TCTGGAGAGA	AGCAGCCGTG	GGCAGAGCCT	GAGGAGATGA	GCGAGGAGAA
,	GTGTGGCTTC	GTTGGCCATG	ACCAGCTGGC	TGGCAGTGAC	GACAGCGAAA	TGGAGGATGA
40	GGCTGAGCCC	CCGGGGGCAC	CCCCTGCACC	CCCGCCCTCC	TATGGGGCCA	CACACAGCAC
	ATTTCAGCCC	CCCAGGCCCC	CACCCCCTGT	CCGGGACTAC	TGACCATGTG	CCTCCCACTG
45	AATGGCAGTT	TCCAGGACCT	CCATTCCACT	CATCTCTGGC	CTGAGTGACA	GTGTCAAGGA
	ACCCTGCTCC	TCTCTGTCCT	GCCTCAGGCC	TAAGAAGCAC	TCTCCCTTGT	TCCCAGTGCT
	GTCAAATCCT	CTTTCCTTCC	CAATTGCCTC	TCAGGGGTAG	TGAAGCTGCA	GACTGACAGT
50	TTCAAGGATA	CCCAAATTCC	CCTAAAGGTT	CCCTCTCCAC	CCGTTCTGCC	TCAGTGGTTT
	CAAATCTCTC	CTTTCAGGGC	TTTATTTGAA	TGGACAGTTC	GACCTCTTAC	TCTCTCTTGT
55	GGTTTTGAGG	CACCCACACC	CCCCGCTTTC	CTTTATCTCC	AGGGACTCTC	AGGCTAACCT

5

TTGAGATCAC TCAGCTCCCA TCTCCTTTCA GAAAAATTCA AGGTCCTCCT CTAGAAGTTT

CAAATCTCTC CCAACTCTGT TCTGCATCTT CCAGATTGGT TTAACCAATT ACTCGTCCCC

GCCATTCCAG GGATTGATTC TCACCAGCGT TTCTGATGGA AAATGGCGGG AATTCCTGCA

GCCCGGGGGA TCCACT

which is SEQ ID NO:1;

10 (b) CGAUAAGCUU GAUAUCGAAU UCCGGACUCU UGCUCGGGCG CCUUAACCCG GCGUUCGGUU CAUCCCGCAG CGCCAGUUCU GCUUACCAAA AGUGGCCCAC UAGGCACUCG CAUUCCACGC 15 CCGGCUCCAC GCCAGCGAGC CGGGCUUCUU ACCCAUUUAA AGUUUGAGAA UAGGUUGAGA UCGUUUCGGC CCCAAGACCU CUAAUCAUUC GCUUUACCGG AUAAAACUGC GUGGCGGGG UGCGUCGGGU CUGCGAGAGC GCCAGCUAUC CUGAGGGAAA CUUCGGAGGG AACCAGCUAC 20 UAGAUGGUUC GAUUAGUCUU UCGCCCCUAU ACCCAGGUCG GACGACCGAU UUGCACGUCA GGACCGCUAC GGACCUCCAC CAGAGUUUCC UCUGGCUUCG CCCUGCCCAG GCGAUCGGCG GGGGGACCC GCGGGGUGAC CGGCGGCAGG AGCCGCCACC AUGGAGUUCC GCCAGGAGGA 25 GUUUCGGAAG CUAGCGGGUC GUGCUCUCGG GAAGCUGCAC CGCCUUCUGG AGAAGCGGCA GGAAGGCGCG GAGACGCUGG AGCUGAGUGC GGAUGGGCGC CCGGUGACCA CGCAGACCCG 30 GGACCCGCCG GUGGUGGACU GCACCUGCUU CGCCCCCCU CGCCGCUACA UUAUCGCCAU CAUGAGUGGU CUGGGCUUCU GCAUCAGCUU UGGCAUCCGC UGCAACCUGG GCGUGGCCAU 35 CGUCUCCAUG GUCAAUAACA GCACGACCCA CCGCGGGGGC CACGUGGUGG UGCAGAAAGC CCAGUUCAGC UGGGAUCCAG AGACUGUCGG CCUCAUACAC GGCUCCUUUU UCUGGGGCUA CAUUGUCACU CAGAUUCCAG GAGGAUUUAU CUGUCAAAAA UUUGCAGCCA ACAGAGUUUU 40 CGGCUUUGCU AUUGUGGCAA CAUCCACUCU AAACAUGCUG AUCCCCUCAG CUGCCCGCGU CCACUAUGGC UGUGUCAUCU UCGUGAGGAU CCUGCAGGGG UUGGUAGAGG GGGUCACAUA 45 CCCCGCCUGC CAUGGGAUCU GGAGCAAAUG GGCCCCACCC UUAGAACGGA GUCGCCUGGC GACGACAGCC UUUUGUGGUU CCUAUGCUGG GGCGGUGGUC GCGAUGCCCC UCGCCGGGGU CCUUGUGCAG UACUCAGGAU GGAGCUCUGU UUUCUACGUC UACGGCAGCU UCGGGAUCUU 50 CUGGUACCUG UUCUGGCUGC UCGUCUCCUA CGAGUCCCCC GCGCUGCACC CCAGCAUCUC GGAGGAGGAG CGCAAGUACA UCGAGGACGC CAUCGGAGAG AGCGCGAAAC UCAUGAACCC

CCUCACGAAG UUUAGCACUC CCUGGCGGCG CUUCUUCACG UCUAUGCCAG UCUAUGCCAU CAUCGUGGCC AACUUCUGCC GCAGCUGGAC GUUCUACCUG CUGCUCAUCU CCCAGCCCGA 5 CUACUUCGAA GAAGUGUUCG GCUUCGAGAU CAGCAAGGUA GGCCUGGUGU CCGCGCUGCC CCACCUGGUC AUGACCAUCA UCGUGCCCAU CGGCGGCCAG AUCGCGGACU UCCUGCGGAG CCGCCGCAUC AUGUCCACCA CCAACGUGCG CAAGUUGAUG AACUGCGGAG GCUUCGGCAU 10 GGAAGCCACG CUGCUGUUGG UGGUCGGCUA CUCGCACUCC AAGGGCGUGG CCAUCUCCUU CCUGGUCCUA GCCGUGGGCU UCAGCGGCUU CGCCAUCUCU GGGUUCAACG UGAACCACCU 15 GGACAUAGCC CCGCGCUACG CCAGCAUCCU CAUGGGCAUC UCCAACGGCG UGGGCACACU GUCGGGCAUG GUGUGCCCCA UCAUCGUGGG GGCCAUGACU AAGCACAAGA CUCGGGAGGA GUGGCAGUAC GUGUUCCUAA UUGCCUCCCU GGUGCACUAU GGAGGUGUCA UCUUCUACGG 20 GGUCUUUGCU UCUGGAGAGA AGCAGCCGUG GGCAGAGCCU GAGGAGAUGA GCGAGGAGAA GUGUGGCUUC GUUGGCCAUG ACCAGCUGGC UGGCAGUGAC GACAGCGAAA UGGAGGAUGA 25 GGCUGAGCCC CCGGGGGCAC CCCCUGCACC CCCGCCCUCC UAUGGGGCCA CACACAGCAC AUUUCAGCCC CCCAGGCCCC CACCCCCUGU CCGGGACUAC UGACCAUGUG CCUCCCACUG AAUGGCAGUU UCCAGGACCU CCAUUCCACU CAUCUCUGGC CUGAGUGACA GUGUCAAGGA 30 ACCCUGCUCC UCUCUGUCCU GCCUCAGGCC UAAGAAGCAC UCUCCCUUGU UCCCAGUGCU GUCAAAUCCU CUUUCCUUCC CAAUUGCCUC UCAGGGGUAG UGAAGCUGCA GACUGACAGU 35 UUCAAGGAUA CCCAAAUUCC CCUAAAGGUU CCCUCUCCAC CCGUUCUGCC UCAGUGGUUU CAAAUCUCUC CUUUCAGGGC UUUAUUUGAA UGGACAGUUC GACCUCUUAC UCUCUUGU GGUUUUGAGG CACCCACACC CCCCGCUUUC CUUUAUCUCC AGGGACUCUC AGGCUAACCU 40 UUGAGAUCAC UCAGCUCCCA UCUCCUUUCA GAAAAAUUCA AGGUCCUCCU CUAGAAGUUU CAAAUCUCUC CCAACUCUGU UCUGCAUCUU CCAGAUUGGU UUAACCAAUU ACUCGUCCCC 45 GCCAUUCCAG GGAUUGAUUC UCACCAGCGU UUCUGAUGGA AAAUGGCGGG AAUUCCUGCA GCCCGGGGGA UCCACU

which is SEQ ID NO:3;

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(c) a nucleic acid compound complementary to (a) or(b); and

- (d) a fragment of (a), (b), or (c) that is at least 18 bases in length and which will selectively hybridize to human genomic DNA encoding a human brain Na⁺-dependent inorganic phosphate cotransporter.
- An expression vector capable of producing a human brain sodium-dependent inorganic phosphate
 cortranporter, or a fragment thereof, in a host cell which comprises a nucleic acid compound as claimed in Claim 3 operably linked with regulatory elements necessary for expression of the nucleic acid compound in the host cell.
- 5. An expression vector as claimed in Claim 4 which comprises a nucleic acid compound encompassing nucleotides 461 to 2143 of SEQ ID NO:1, or a sequence complementary to this region.

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6. A method of evaluating the effectiveness of a test compound for the treatment or prevention of a condition associated with an inappropriate stimulation of a human Na⁺⁻ dependent inorganic phosphate cotransporter protein which method comprises:

a) introducing into a mammalian host cell an expression vector comprising DNA encoding a human hBNPI protein as claimed in either one of Claims 2 or 3;

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b) culturing said host cell under conditions such that the human hBNPI protein is expressed;

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c) exposing said host cell expressing the human hBNPI protein to a test compound; and

- d) measuring the change in a physiological condition known to be influenced by the binding of native ligand to the human hBNPI protein relative to a control in which the transfected host cell is exposed to native ligand.
- 7. A method of evaluating the effectiveness of a test compound for the treatment or prevention of a condition associated with an inappropriate stimulation of a human Na⁺-dependent inorganic phosphate cotransporter protein compounds which method comprises:

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a) introducing into a mammalian host cell an expression vector comprising DNA encoding a human Na+-dependent inorganic phosphate cotransporter protein as claimed in either one of Claims 2 or 3;

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b) culturing said host cell under conditions such that the human Na⁺-dependent inorganic phosphate cotransporter protein is expressed;

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c) exposing said host cell expressing the human Na⁺-dependent inorganic phosphate cotransporter protein to a test compound;

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d) exposing said host cell expressing the Na+-dependent inorganic phosphate cotransporter protein to inorganic phosphate simultaneously with or following the exposure to the test compound; and

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e) measuring the change in inorganic phosphate uptake relative to a control in

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which the transfected host cell is exposed to only inorganic phosphate.

INTERNATIONAL SEARCH REPORT

International application No. PCT/US96/05792

A. CLASSIFICATION OF SUBJECT MATTER IPC(6) :Please See Extra Sheet.							
US CL :Please See Extra Sheet.							
According to International Patent Classification (IPC) or to both national classification and IPC							
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1	documentation searched (classification system follow	•					
0.5. :	435/7.1, 69.1, 252.3, 240.1, 320.1; 530/300, 350;	436/501; 536/23.1					
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched							
Electronic	data base consulted during the international search (n	ame of data base and, where practicable	, search terms used)				
Please See Extra Sheet.							
C. DOG	CUMENTS CONSIDERED TO BE RELEVANT						
Category*	Citation of document, with indication, where a	ppropriate, of the relevant passages	Relevant to claim No.				
X .	Ni et al. Cloning and expression of a cDNA encoding a brain- specific Na-dependent inorganic phosphate cotransporter. Proc. Natl. Acad. Sci. USA. June 1994, Vol. 91, pages 6, 7						
Υ							
•	5607-5611, especially pages 560		<i>0, 7</i>				
Y	Chong et al. Molecular Cloning	of the cDNA Encoding a	1-7				
•	Human Renal Sodium Phosphate		1-7				
	Assignment to Chromosome 6						
	November 1993, Vol. 18, pages 3	55-359, especically pages					
355-357.							
A, P	Li et al. Molecular cloning of two rat Na/Pi cotransporters: 1-7						
	evidence for differential tissue expression of transcripts.						
	Cellular and Molecular Biology Re	search. March 1996, Vol.					
	5, pages 451-460.						
	•						
X Further documents are listed in the continuation of Box C. See patent family annex.							
 Special categories of cited documents: T hater document published after the international filing date or priority date and not in conflict with the application but cited to understand the 							
to	"A" document defining the general state of the art which is not considered principle or theory underlying the invention to be part of particular relevance						
	CONTRICTED BOAST OF CHANGE AS CONTRICTED TO DIAGRACIE IN PRACTICAL SECTION OF THE CONTRICTED TO DIAGRACIE AND DIAG						
cita	ed to establish the publication date of another citation or other scial reason (as specified)	"Y" document of particular relevance; the					
	current referring to an oral disclosure, use, exhibition or other	considered to involve an investive combined with one or more other such being obvious to a person skilled in th	documents, such combination				
	cument published prior to the international filing date but later than priority date claimed	*&* document member of the same patent	facuity				
Date of the actual completion of the international search Date of mailing of the international search report							
03 JUNE	1996	15 JUL 1996					
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks		Authorized officer					
Box PCT	a, D.C. 20231	KENNETH A. SORENSEN					
Facsimile No. (703) 305-3230		Telephone No. (703) 308-0196					

INTERNATIONAL SEARCH REPORT

• 65 A ,

International application No. PCT/US96/05792

C (Continua	tion). DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
x	Ni et al. Cloning and expression of a novel cDNA encoding a brain specific Na-dependent inorganic phosphate cotransporter. In: Abstracts of the Society for Neuroscience, 24th Annual Meeting. Volume 20, 1994, Abstract 382.4, page 925.	1-5
Y	Collins et al. Molecular cloning, functional expression, tissue distribution, and in situ hybridization of the renal sodium phosphate (Na/Pi) transporter in the control and hypophospatemic mouse. FASEB Journal, August 1994, Vol. 8, pages 862-868, especially pages 862-865.	1-7
Y	Magagnin et al. Expression cloning of human and rat renal cortex Na/Pi cotransport. Proc. Natl. Acad. Sci. USA. July 1993, Vol. 90, pages 5979-5983, especially pages 5981-5983.	1-7
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INTERNATIONAL SEARCH REPORT

International application No. PCT/US96/05792

A.	CLASSIFICATION	OF	SUBJECT	MATTER:
IP	C (6):			

G01N 33/566; C12P 21/06; C12N 1/20, 15/00; A61K 38/00; C07K 1/00; C07H 21/02

A. CLASSIFICATION OF SUBJECT MATTER: US CL :

435/7.1, 69.1, 252.3, 240.1, 320.1; 530/300, 350; 436/501; 536/23.1

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

STN/MEDLINE, EMBASE, BIOSIS, CONFSCI, DISSABS, WPIDS, PATOSEP JICST-EPLUS, APS search terms: , human brain sodium dependent inorganic phosphate co-transporter, protein, animo acid sequence, cDNA, recombinant, hBNPI, synonyms and authors

Form PCT/ISA/210 (extra sheet)(July 1992)*